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(54) Title: TRANSDUCTION OF RECOMBINASES FOR INDUCIBLE GENE TARGETING

(57) Abstract: The present invention provides the use of a fusion protein comprising a site-specific DNA recombinase domain and a protein transduction domain for preparing an agent for inducing target gene alteration in a living organism or in cultured cells, suitable fusion proteins and a method for the production of said fusion proteins.

Transduction of recombinases for inducible gene targeting

The present invention provides the use of a fusion protein comprising a site-specific DNA recombinase domain and a protein transduction domain for preparing an agent for inducing target gene alteration in a living organism or in cultured cells, suitable fusion proteins and a method for the production of said fusion proteins.

Background

For some years targeted mutagenesis in totipotent mouse embryonic stem (ES) cells has been used to inactivate genes, for which cloned sequences were available (Capecchi, Trends in Genetics 5, 70 - 76 (1989)). Since ES cells can pass mutations induced *in vitro* to transgenic offspring *in vivo*, it is possible to analyze the consequences of gene disruption in the context of the entire organism. Thus, numerous mouse strains with functionally inactivated genes ("knock out mice") have been created by this technology and utilized to study the biological function of a variety of genes.

A refined method of targeted mutagenesis, referred to as conditional mutagenesis, employs a site-specific recombination system (e.g. Cre/loxP or Flp/frt – Sauer and Henderson, N. Proc. Natl. Acad. Sci. USA 85, 5166-5170 (1988); Senecoff et al., J. Mol. Biol., 201, 405 - 421 (1988)) which enables a temporally and/or spatially restricted alteration of target genes (Rajewsky et al., J. Clin. Invest., 98, 600 - 603 (1996)). The creation of conditional mouse mutants requires the generation of two mouse strains, i.e. the recombinase recognition strain and the recombinase expressing strain. The recombinase recognition strain is generated by homologous recombination in ES cells as described above except that the targeted

exon(s) is (are) flanked by two recombinase recognition sequences (hereinafter "RRS"; e.g. loxP or *frt*). The type of recombination event mediated by the recombinase depends on the disposition of the RRS, with deletions, inversions, translocations and integrations being possible (Torres and Kühn, Oxford University Press, Oxford, New York (1997)). By placing the RRS into introns, an interference with gene expression before recombination can be avoided. The recombinase expressing strain contains a recombinase transgene (e.g. Cre, Fip) whose expression is either restricted to certain cells and tissues or is inducible by external agents. Crossing of the recombinase recognition strain with the recombinase expressing strain recombines the RRS-flanked exons from the doubly transgenic offspring in a prespecified temporally and/or spatially restricted manner. Thus, the method allows the temporal analysis of gene function in particular cells and tissues of otherwise widely expressed genes. Moreover, it enables the analysis of gene function in the adult organism by circumventing embryonic lethality which is frequently the consequence of gene mutation. For pharmaceutical research, aiming to validate the utility of genes and their products as targets for drug development, inducible mutations provide an excellent genetic tool. However, the current systems for inducible recombinase expression in transgenic animals suffer from a certain degree of leakiness in the absence of the inducer (Kühn et al., *Science* 269(5229):1427-9 (1995); Schwenk et al., *Nucleic Acids Res.*; 26(6):1427-32 (1998)). Furthermore, the generation of conditional mutants is a time consuming and labor intensive procedure, since the recombinase recognition strain and the recombinase expressing strain have to be breed at least over two generations in order to obtain animals carrying both, the recombinase transgene and two copies of the RRS-flanked target gene sequence.

Protein transduction domains (hereinafter shortly referred to as "PTD") that have the ability to cross cell membranes were identified, e.g. in the

Antennapedia protein from *Drosophila* (Vives et al., J. Biol. Chem., 272(25):16010-7 (1997)), Kaposi fibroblast growth factor (Kaposi FGF; Lin et al., J. Biol. Chem. 270: 14255-58 (1995)), VP22 from HSV (Elliott and O'Hare, Cell, 88(2):223-33 (1997)) and TAT from HIV (Green and Loewenstein, Cell, 55(6):1179-88 (1988); Frankel and Pabo, Cell, 55(6):1189-93 (1988)). WO 99/29721 moreover mentions TAT mutants having an enhanced activity as compared to the wild-type peptide.

Fusion of PTDs to heterologous proteins conferred the ability to transduce into cultured cells (Fawell et al., Proc. Natl. Acad. Sci. USA, 91(2):664-8 (1994); Elliott and O'Hare (1997), Phelan et al., Nature Biotech. 16; 440-443 (1998) and Dilber et al., Gene Ther., 6(1):12-21 (1999)). Dalby and Bennett showed that a fusion protein consisting of VP22 and functional Flp recombinase translocated between cells in culture (from COS-1 cells transfected with VP22-Flp to CHO cells carrying Flp recognition sites (FRT sites); see Dalby and Bennett, Invitrogen, Expressions 6.2, page 13 (1999)). Further WO 99/11809 mentions a fusion protein Antp-Cre and emphasizes that it may be used to deliver the Cre into the cell which recombines inside the cell nucleus. It is mentioned that the fusion protein is suitable for manipulating genomic DNA at precise locations in a temporal regulated manner.

Furthermore, a recent report demonstrated that the β -galactosidase protein fused to the 11 amino acids PTD from the HIV TAT protein can infiltrate all tissues of living mice reaching every single cell (Schwarze et al., Science, 285(5433):1569-72 (1999)). Finally, WO 99/60142 discloses vector constructs for gene therapy carrying a tumor cell sensitizing gene, a sensitizing gene expression regulatory system, a control gene and a control gene expression regulatory system, wherein the control gene can be a fusion gene consisting of a recombinase (viz. Cre or Flp) and a trafficking protein (viz. VP22).

With regard to the fusion protein Antp-Cre of WO 99/11809, it is however, general knowledge in the art that the Antennapedia PTD is not a generally applicable transducing protein, namely it has only a limited activity with proteins having more than 100 amino acid residues (Derossi et al., Trends Cell Biol. 8: 84-87, 1998). In view of the limited transducing activity of the Antp PTD and the size of the generally known recombinases (ranging from about 200 to about 600 amino acid residues), it was desirable to provide a more potent system for the transduction of recombinases. It was, however, not clear for a person skilled in the art whether PTDs would be effective at all with recombinases for the following reasons:

- (i) only a single example of PTD-mediated delivery of proteins (above 100 amino acid residues) *in vivo* has been reported so far (Schwarze et al., Science, 285(5433):1569-72 (1999); Fawell et al., PNAS, 91: 664-68 (1994); both references describing the TAT-mediated transduction of β -galactosidase in mice);
- (ii) It is known that - due to defolding and refolding processes - the transduction of native proteins into cells may result in a significant loss of protein activity (e.g., as described for TAT-GFP; Schwarze et al, Trends Cell Biol. 10: 290-95 (2000));
- (iii) neither the number of protein molecules that can be transferred into a cell by a given translocation domain has been systematically determined, nor the number of Cre molecules in the cell nucleus that is required for efficient recombination;
- (iv) the delivery of active proteins requires unfolding- and proper refolding which is unpredictable for a given protein (Bonifaci et al., AIDS 9: 995-1000 1995); and
- (v) the mechanism by which protein transduction domains facilitate protein transduction is unknown and several findings have been published that rule out classical receptor-, transporter-, endosome- or endocytosis-mediated processes in the transduction of Ant, TAT and VP22 (G. Elliott, P. O'Hare, Cell 88, 223-233 (1997); D.A. Mann, A.D. Frankel, EMBO. J. 10,

1733-1739 (1991); D. Derossi et al., J. Biol. Chem. 269, 10444-10450 (1994); D. Derossi et al., J. Biol. Chem. 271, 18188-18193 (1996); E. Vives et al., J. Biol. Chem. 272, 16010-16017 (1997)).

Moreover, there was still the need for a generally applicable method where the genetic manipulation can be performed in both, endogenous genes and transgenes.

Summary of the Invention

It was found that site-specific DNA recombinase proteins can be translocated into cells of a living organism when fused to specific protein transduction domains, namely transduction domains being derived from the VP22 protein of HSV or from the TAT protein of HIV. Thus, whenever a gene mutation is desired, recombination is induced upon the injection of the appropriate site-specific recombinase fused to a transduction domain into such a living organism (provided, however, that said organism carries at least one appropriate RRS integrated in the genome).

The present invention thus provides

- (1) the use of a fusion protein comprising
 - (a) a site-specific DNA recombinase domain and
 - (b) a protein transduction domain (PTD)

for preparing an agent for inducing target gene alterations in a living organism or cell culture, wherein said living organism carries at least one or more recognition sites for said site-specific DNA recombinase integrated in its genome;

- (2) a method for inducing gene alterations in a living organism which comprises administering to said living organism a fusion protein comprising a site-specific DNA recombinase domain and a PTD as defined in (1) above, wherein said living organism carries at least one or more

recognition sites for said site-specific DNA recombinase integrated in its genome;

(3) a fusion protein comprising

(a) a site-specific DNA recombinase domain and

(b) a PTD being derived from the VP22 protein of HSV or from the TAT protein of HIV

provided that when the site-specific DNA recombinase domain is wild-type Cre or Flp then the PTD is not the full length VP22 PTD of HSV (i.e., the fusion protein is not identical to the fusion protein of Dalby and Bennett, Invitrogen, Expressions 6.2, page 13 (1999) and of WO 99/60142);

(4) a DNA sequence coding for the fusion protein of (3) above;

(5) a vector comprising the DNA sequence as defined in (4) above;

(6) a host cell transformed with the vector of (5) above and/or comprising the DNA of (4) above;

(7) a method for producing the fusion protein of (1) above which comprises culturing the transformed host cell of (6) above and isolating the fusion protein; and

(8) an injectable composition comprising the fusion protein as defined in (1) or (3) above.

The invention is further illustrated by the appended Figures and is explained in detail below.

Description of the Figures

Fig. 1: Generation of induced mouse mutants using purified fusion proteins.

A: Expression of the fusion protein consisting of the site-specific DNA recombinase (e.g. Cre) and the protein transduction domain (e.g. the HIV derived TAT peptide) in prokaryotic or eukaryotic cells.

B: Extraction and purification of the expressed fusion protein (e.g. as described in Nagahara et al., Nat. Med. 4 (12):1449-52 (1998)).

C: Injection of the purified fusion protein into mice carrying the RRS-flanked target sequence.

D: Analysis of the pattern of induced target gene recombination and the resulting phenotype.

Triangle: RRS.

Fig. 2: Scheme of the bacterial expression vector pT7-TACS (SEQ ID NO:16). The coding region of the 11 amino acid protein transduction domain of HIV TAT protein is fused to the N-terminus of the Cre recombinase protein sequence. The 10-amino-acid strep tag and the protease factor Xa recognition sequence are fused to the C-terminus. The T7 promoter permits expression of TAT-Cre protein in *E. coli*.

Fig. 3: Detection of purified TAT-Cre protein by Coomassie staining and Western blot analysis.

A: Coomassie stained SDS-PAGE gel. Lane 1: 10 kDa ladder (Life Technologies, Cat. No.: 10064-012), 2: 1000 ng BSA, 3: 750 ng BSA, 4: 500 ng BSA, 5: 100 ng BSA, 6: 50 ng BSA, 7: 5 μ l TAT-Cre, 8: 1 μ l TAT-Cre in Bicine buffer.

B: Western blot analysis using an alkaline phosphatase-conjugated anti-strep tag antibody (IBA, Cat. No: 2-1503-001). Lane 1: MultiMark (Invitrogen, Cat. No.: LC5725), 2: 7 μ l TAT-Cre, 3: 5 μ l TAT-Cre, 4: 2,5 μ l TAT-Cre, 5: 1,25 μ l TAT-Cre in Bicine buffer.

Fig. 4: X-Gal staining of M5Pax8 cells treated with TAT-Cre protein. M5Pax8 fibroblasts were treated for 18 h with 3,5 (A), 6,9 (B) and 13,8 μ g/ml TAT-Cre protein (C) in serum-free medium. Four days after treatment, cells were fixed and stained with X-Gal.

Fig. 5: Measurement of β -galactosidase activity in cell lysates. M5Pax8 fibroblasts were treated for 18 h with increasing concentrations of TAT-Cre, as indicated, or transiently transfected with either expression vectors

for Cre (pCMV-I-Cre-pA, see SEQ ID NO:29) or β -galactosidase (pCMV-I- β -pA, see SEQ ID NO:30). Four days after treatment, cells were lysed and the β -galactosidase activities were determined.

Fig. 6: PCR detection of TAT-Cre mediated recombination in mice.

A: PCR-analysis of genomic DNA from duodenum (lane 2), liver (3), kidney (4), spleen (5), muscle (6), lung (7), tail (8) and brain (9) of a *pln13* mouse treated three times with intraperitoneal injections of 75 μ g TAT Cre protein at two-day-intervals. Deletion of the loxP-flanked DNA segment is indicated by the presence of the about 400 bp fragment. Lane 1: 1-kb-ladder (Life Technologies).

B: PCR strategy to detect Cre-mediated deletion of the loxP-flanked DNA segment. Arrows indicate the positions of the primers.

C: PCR-analysis of genomic DNA from spleen of a *pln13* mouse treated three times with intraperitoneal injections of 75 μ g TAT Cre protein at two-day-intervals (lane 4). To confirm the presence of the BamH I restriction site, the PCR product was digested with BamH I which produces two diagnostic fragments of about 190 and about 210 bp (5). As a control, tail DNA from untreated mice carrying the loxP-flanked (lane 2) and the detected *pln13* allele (3) was subjected to PCR amplification. Lane 1: 100 bp ladder (Life Technologies), lane 6: 1 kb ladder (Life Technologies).

Fig. 7: Scheme of the bacterial expression vectors pT7-VPCS (SEQ ID NO:17) and pCRT7- Δ VPCS (SEQ ID NO:15). The coding region of the 301 amino acid protein transduction domain of HSV VP22 protein (A) or the truncated 143 amino acid Δ VP22 domain (B) is fused to the N-terminus of the Cre recombinase protein sequence. The 10-amino-acid strep tag and the protease factor Xa recognition sequence are fused to the C-terminus. The T7 promoter allows the expression of VP22-Cre and Δ VP22-Cre fusion proteins in *E. coli*. The sequence in pCRT7- Δ VPCS encoding the 15 amino

acid N-terminal leader sequence is used for enhanced protein stability (Invitrogen).

Fig. 8: Detection of the purified VP22-Cre and Δ VP22-Cre fusion proteins by Coomassie staining and Western blot analysis.

A: Detection of VP22-Cre protein in a Coomassie-stained SDS-PAGE gel. Lane 1: 10 kDa ladder, 2: 1000 ng BSA, 3: 500 ng BSA, 4: 100 ng BSA, 5: inclusion body protein extract before chromatography, 6: unbound protein, 7: fraction 17, 8: fraction 18, 9: fraction 19, 10: fraction 20. The position of the 75 kDa VP22-Cre protein is indicated by the arrow head.

B: Detection of VP22-Cre protein by Western blot analysis using an alkaline phosphatase-conjugated anti-strep tag antibody (IBA, Cat. No.: 2-1503-001). Lane 1: MultiMark (Invitrogen), 2: inclusion body protein extract before chromatography, 3: unbound protein, 4: fraction 10, 5: fraction 11, 6: fraction 16, 7: fraction 17, 8: fraction 18, 9: fraction 19, 10: fraction 20.

C: Detection of Δ VP22-Cre protein in a Coomassie-stained SDS-PAGE gel. Lane 1: 10 kDa ladder, 2: inclusion body protein extract before chromatography, 3: unbound protein, 4: fraction 1, 5: fraction 8, 6: fraction 9, 7: fraction 15, 8: 100 ng BSA, 9: 500 ng BSA, 10: 1000 ng BSA. The position of the 60 kDa Δ VP22-Cre protein is indicated by the arrow head.

D: Detection of Δ VP22-Cre protein by Western blot analysis using an alkaline phosphatase-conjugated anti-strep tag antibody (IBA, Cat. No.: 2-1503-001). Lane 1: MultiMark (Invitrogen), 2: inclusion body protein extract before chromatography, 3: unbound protein, 4: fraction 4, 5: fraction 8, 6: fraction 10, 7: fraction 12, 8: soluble protein extract before chromatography, 9: unbound protein, 10: fraction 7.

Fig. 9: X-Gal staining of M5Pax8 cells treated with VP22-Cre and Δ VP22-Cre fusion proteins. M5Pax8 fibroblasts were treated for 18 h with either

Bicine buffer (A), 0.5 µg/ml VP22-Cre (B) or 3.75 µg/ml ΔVP22-Cre (C) in serum-free medium. Four days after treatment, cells were fixed and stained with X-Gal.

Fig. 10: Measurement of β-galactosidase activity in cell lysates. M5Pax8 fibroblasts were treated for 18 h with VP22-Cre, ΔVP22-Cre or Bicine buffer alone, as indicated or transiently transfected with expression vectors for Cre (pCMV-I-Cre-pA, see SEQ ID NO:29) or β-galactosidase (pCMV-I-β-pA, see SEQ ID NO:30). Four days after treatment, cells were lysed and the β-galactosidase activities were determined.

Fig. 11: PCR detection of Cre mediated recombination in cells treated with VP22-Cre and ΔVP22-Cre fusion proteins shown in SEQ ID NOs: 21 and 14, respectively).

A: PCR-analysis of genomic DNA isolated from M5Pax8 fibroblasts. Cells were transiently transfected with a Cre expression vector (lane 2) or treated for 18 h with either buffer alone (lane 3), 7.5 µg/ml VP22-Cre (4, 5) or 15 µg/ml ΔVP22-Cre (6, 7) in serum-free medium. Four days after treatment, genomic DNA was extracted and subjected to PCR amplification. Deletion of the loxP-flanked DNA segment is indicated by the presence of the 226 bp DNA fragment. To confirm the presence of the Nco I restriction site in the recombined allele, the PCR products were digested with Nco I which produces two diagnostic fragments of 85bp and 141bp (lanes 5 and 7). Lane 1: 100 bp ladder (Life Technologies), lane 8: 1 kb ladder (Life Technologies).

B: PCR strategy to detect Cre-mediated deletion of the loxP-flanked DNA segment. Arrows indicate the positions of the primers.

Detailed Description of the Invention

The expression "target sequences" according to the present invention means all kind of sequences which may be mutated (viz. deleted,

translocated, integrated and/or inverted) by the action of the recombinase. The number of RRS in the target sequence depends on the kind of mutation to be performed by the recombinase. For most of the mutations (especially for deletions and inversions) two RRS are required which are flanking the sequence to be mutated (deleted or inverted). For some kinds of integrations only one RRS may be necessary within the target sequence.

The "living organisms" according to the present invention are multi-cell organisms and can be vertebrates such as mammals (e.g., rodents such as mice or rats) or non-mammals (e.g., fish) or can be invertebrates such as insects or worms, or can be plants (higher plants, algae or fungi). Most preferred living organisms are mice and fish.

"Cell culture" according to the present invention include cells isolated from the above defined living organism and cultured *in vitro*. These cells can be transformed (immortalized) or untransformed (directly derived from the living organism; primary cell culture).

The site-specific DNA recombinase domain within the fusion protein of the invention of the present application is preferably selected from a recombinase protein derived from Cre, FLP, ϕ C31 recombinase (Thorpe and Smith, Proc. Natl. Acad. Sci, USA, vol. 95, 5505-5510 (1998)), $\gamma\delta$ resolvase (Schwickardi and Dröge, FEBS letters 471:147-150 (2000) and R recombinase (Araki et al., J. Mol. Biol., 182, 191-203 (1985)). The preferred recombinases are Cre and mutants thereof (preferably the Cre variant of aa 15 to 357 of SEQ ID NO: 2 or aa 325-667 of SEQ ID NO: 6) and FLP and variants thereof including FLPe (preferably the FLP variant of aa 15 to 437 of SEQ ID NO: 4 or aa 325 to 747 of SEQ ID NO: 8).

The protein transduction domain according to the present invention includes, but is not limited to, the PTDs mentioned in Background of the Invention. The PTD preferably is derived from the VP22 protein of HSV or from the TAT protein of HIV. Suitable TAT proteins include, but are not limited to, proteins comprising (i) the amino acid sequence shown in SEQ ID NO: 10 and mutant thereof such as

(ii) proteins comprising the amino acid

AGRKKRRQRRR (SEQ ID NO:22)

YARKARRQARR (SEQ ID NO:23)

YARAAARQARA (SEQ ID NO:24)

YARAARRAARR (SEQ ID NO:25)

YARAARRAARA (SEQ ID NO:26)

YARRRRRRRRR (SEQ ID NO:27)

YAAARRRRRRR (SEQ ID NO:28)

as known from WO 99/29721. Preferred are transduction domains consisting of the TAT proteins (i) and (ii) above.

Suitable VP22 proteins include, but are not limited to, the wild-type VP22 protein, i.e., a protein comprising amino acids 1 to 302 of SEQ ID No:21, and truncated forms thereof. Truncated VP22 proteins in accordance with the present invention can be those lacking 1 to 158 amino acid residues at their N-terminal end. The most preferred VP22 protein is the truncated VP22 PTD comprising amino acid residues 16 to 157 of SEQ ID NO:14.

The fusion of the two domains of the fusion protein can occur at any possible position, i.e., the protein transduction domain can be fused to the N- or C-terminal of the site-specific DNA recombinase or can be fused to active sites within the site-specific DNA recombinase. Preferably the protein transduction domain is fused to the N-terminal of the site-specific DNA recombinase domain.

The protein transduction domain can be fused to the site-specific DNA recombinase either through a direct chemical bond or through a linker molecule. Such linker molecule can be any bivalent chemical structure capable of linking the two domains. The preferred linker molecule according to the present invention is a short peptide, e.g., having 1 to 20, preferably 1 to 10, amino acid residues. Specifically preferred short peptides are essentially consisting of Gly, Ala and/or Leu.

The fusion protein of the invention of the present application may further comprise other functional sequences such as secretion conferring signals, nuclear localisation signals and/or signals conferring protein stabilisation.

In case the fusion protein comprises a protein transduction domain derived from the TAT protein of HIV, the DNA sequence coding for said fusion protein preferably comprises the sequence

5' TAC GGC CGC AAG AAG CGC CGC CAA CGC CGC CGC 3'.

Such a preferred DNA sequence is for instance shown in SEQ ID NO: 11. In said sequence the 3' terminal codon ggc codes for the linker Gly. The DNA sequence of a suitable recombinase may be directly attached to said codon ggc.

The fusion protein can be obtained by the following steps:

1. Fusion of the recombinase coding region (e.g. encoding Cre: see amino acids 15 to 357 of SEQ ID NO: 2) with the sequence conferring protein translocation (e.g. the sequence encoding the TAT peptide YGRKKRRQRRR, SEQ ID NO: 10) using standard cloning protocols (Maniatis et al., Cold Spring Harbor Laboratory, New York (1989)) or chemical synthesis.

2. Generation of a construct for the expression of the fusion protein in prokaryotic or eukaryotic cells, e.g. in *E. coli* DH5a (Hanahan, J. Mol. Biol.;166(4):557-80 (1983)) using the QIAexpress pQE vector (Qiagen, Hilden).
3. Expression of the above mentioned fusion protein in prokaryotic or eukaryotic cells, e.g. in *E. coli* DH5a (Hanahan, 1983)
4. Extraction and purification of the above mentioned fusion protein e.g. as described in Nagahara et al., Nat. Med., 4(12):1449-52 (1998).

In an experiment it was shown that TAT-mediated delivery of active Cre protein works with sufficient efficacy to facilitate inducible gene targeting both in cell lines and living organisms. In this experiment a vector for the expression of a TAT-Cre fusion protein in *E. coli* was constructed, TAT-Cre protein was expressed in *E. coli* and purified from bacterial lysates. To test the activity of the TAT-Cre protein *in vitro*, a reporter cell line that contains a loxP-containing reporter construct was used. This reporter, when recombined by Cre recombinase, allows the expression of a β -galactosidase gene. Further, a transgenic mouse strain carrying a loxP-flanked target was used to invest the activity of the TAT-Cre protein *in vivo*.

In a second experiment it was shown that VP22-mediated delivery of active Cre protein works with sufficient efficacy to facilitate inducible gene targeting. In this experiment Bacterial expression vectors were constructed for the production of VP22-Cre fusion proteins in *E. coli*. The activity of purified VP22-Cre proteins were tested using a reporter fibroblast cell line containing a loxP-flanked reporter construct.

Thus, the injection of the purified fusion protein of the present invention into a living organism (e.g., a mouse) carrying a gene comprising the RRS-flanked target sequence (e.g., in an amount of 1 to 200, preferably 5

to 50 µg per g body weight). To demonstrate the feasibility of the invention, a reporter mouse strain carrying an RRS-flanked cassette was used (Thorey et al., Mol. Cell Biol., 18(10):6164 (1998)).

Analysis is achieved by determining the pattern of induced target gene recombination (e.g. through PCR analysis, Southern blot analysis or X-Gal staining on tissue sections; Maniatis et al., 1989; Gossler and Zachgo, Joyner AL (Ed.), Oxford University Press, Oxford, New York (1993)).

The procedure's advantages over current technology are as follows:

- (i) The absence of background recombination before administration of the fusion protein.
- (ii) The reduction of time and resources which are necessary to combine the recombinase transgene and two copies of the RRS-flanked target gene by conventional breeding.

In experiments it was shown the following: (a) With a suitable vector for the expression of a TAT-Cre fusion protein, a TAT-Cre fusion protein was expressed in *E. coli* and purified from bacterial lysates.

(b) A reporter cell line containing a loxP-containing reporter construct was used to test the activity of the TAT-Cre protein *in vitro*. This reporter, when recombined by Cre recombinase, allows the expression of a β -galactosidase gene.

(c) A transgenic mouse strain carrying a loxP-flanked target was used to invest the activity of the TAT-Cre protein *in vivo*.

These experiments demonstrate that TAT-mediated delivery of active Cre protein works with sufficient efficacy to facilitate inducible gene targeting both in cell lines and living organisms.

Furthermore, bacterial expression vectors were constructed for the production of VP22-Cre fusion proteins in *E. coli*. The activity of purified VP22-Cre proteins were tested using a reporter fibroblast cell line containing a loxP-flanked reporter construct. These experiments demonstrate that VP22-mediated delivery of active Cre protein works with sufficient efficacy to facilitate inducible gene targeting.

The invention is further illustrated by the following, non-limitative examples.

Examples

Materials and Methods

Construction of pT7-TACS: The TAT-Cre coding region was generated by PCR using Advantage-HF PCR Kit (Clontech), 20 pmol of the primers TATcre sense (5'-atg cca tgg gct acg gcc gca aga agc gcc gcc aac gcc gcc gcg gca tgt cca att tac tga ccg tac acc-3'; SEQ ID NO:31) and TATcre antisense (5'-ttt cgg atc cgc cgc ata acc agt g-3'; SEQ ID NO:32) and 10 ng pCMV-I-Cre-pA (see SEQ ID NO:29) as template. The PCR reaction was performed using the following cycle profile: 2' 94 °C, 4 x (30" 94 °C min, 30" 50 °C, 1' 72 °C), 12 x (30" 94 °C min, 30" 55 °C, 1' 72 °C) and 10' 72 °C. The resulting PCR fragment was digested with Nco I and BamH I, treated with Klenow enzyme and ligated into the plasmid pBSII KS+ which had been opened with restriction enzyme BamH I, treated with Klenow and dephosphorylated with calf intestinal phosphatase. The resulting plasmid pBS TAT-5'cre was verified by DNA sequencing. The Plasmid pCMV-I-Cre-pA (SEQ ID NO:29) was digested with Age I and Sal I which released a 1,036 kb fragment containing the 3' part of the Cre coding region. This fragment was ligated into the plasmid pBS TAT-5'cre which had been opened with Age I and Sal I.

10 ng pBS-TATCre was subjected to PCR amplification using 20 pmol of primers FPA001 (5'-tat atc tag acc atg ggc tac ggc cgc aag aag c-3'; SEQ ID NO:33) and FPA002 (5'-gct acc acg acc ttc gat acc atc gcc atc ttc cag cag gcg c-3'; SEQ ID NO:34). PCR was performed using 2,5 U Platinum Pfx DNA polymerase (Gibco BRL) and 2 x Enhancer Solution (Gibco BRL) according to the manufacturers protocol. The following cycle profile was used: 2' 94 °C, 25 x (30" 94 °C min, 15" 54,6 °C, 2'30" 68 °C). The amplified PCR fragment was purified using GFX columns (Amersham Pharmacia), digested with Xba I and ligated into the plasmid pASK57 (Skerra and Arne, Gene 151: 131-135 (1994)) which had been opened with restriction enzymes Xba I and Eco 47 III and dephosphorylated with calf intestinal phosphatase. The resulting plasmid pASK75-TACS was digested with restriction enzymes Nco I and Hind III which released a 1,1 kb fragment. The fragment was subsequently ligated into the plasmid pT7-7 (Studier and Moffatt, J. Mol. Biol. 189: 113-130 (1986)) which had been opened with restriction enzymes Nco I and Hind III and dephosphorylated with calf intestinal phosphatase resulting in the plasmid pT7-TACS (SEQ ID NO:16).

Construction of pT7-VPCS: The Cre coding region was generated by PCR using Advantage-HF PCR Kit (Clontech), 20 pmol of the primers VP22cre sense (5'-taa cta gcg gcc gca tgt cca att tac tga ccg tac ac-3'; SEQ ID NO:35) and VP22cre antisense (5'-tcg agc ggc cgc cat cgc cat ctt cca gca ggc g-3'; SEQ ID NO:36) and 10 ng pgkcre-pA (SEQ ID NO:40) as template. The PCR reaction was performed using the following cycle profile: 2' 94 °C, 5 x (30" 94 °C, 30" 50 °C, 2' 72 °C), 15 x (30" 94 °C, 30" 55 °C, 2' 72 °C) and 10' 72 °C. The resulting PCR fragment was digested with Not I and ligated into the plasmid pVP22/Myc-His (Invitrogen), which had been opened with restriction enzyme NotI, dephosphorylated with calf intestinal phosphatase. The resulting plasmid pVP22-cre myc/His was verified by DNA sequencing.

10 ng pVP22-cre myc/His was subjected to PCR amplification using 20 pmol of primers FPA004 (5'-tat atc tag aca tat gac ctc tcg ccg ctc cg-3'; SEQ ID NO:37) and FPA002 (SEQ ID NO:34). PCR was performed using 2,5 U Platinum Pfx DNA polymerase (Gibco BRL) and 2 x Enhancer Solution (Gibco BRL) according to the manufacturers protocol. The following cycle profile was used: 2' 94 °C, 25 x (30" 94 °C min, 15" 54,6 °C, 2'30" 68 °C). The amplified PCR fragment was purified using GFX columns (Amersham Pharmacia), digested with Xba I and ligated into the plasmid pASK57 (Skerra and Arne, Gene 151: 131-135 (1994)) which had been opened with restriction enzymes Xba I and Eco 47 III and dephosphorylated with calf intestinal phosphatase. The resulting plasmid pASK75-VPCS was digested with restriction enzymes Nde I and Hind III which released a 2,0 kb fragment. The fragment was subsequently ligated into the plasmid pT7-7 (Studier and Moffatt, J. Mol. Biol. 189: 113-130 (1986)) which had been opened with restriction enzymes Nde I and Hind III and dephosphorylated with calf intestinal phosphatase resulting in the plasmid pT7-VPCS (SEQ ID NO:17).

Construction of pCRT7-ΔVPCS: The ΔVP22-Cre coding region was generated by PCR using Platinum Pfx DNA polymerase (Life Technologies), 20 pmol of the primers FPA007 (5'-ttc cga aga cga cga aac acc-3'; SEQ ID NO:38) and FPA008 (5'-tat att cga agc tta tta acc acc gaa ctg cg-3'; SEQ ID NO:39) and 30 ng pT7-VPCS (SEQ ID NO:17) as template. The PCR reaction was performed using the following cycle profile: 2' 94 °C, 25 x (30" 94 °C, 30" 61 °C, 2'30" 68 °C) and 7' 68 °C. The resulting 1,8 kb PCR fragment was digested with Nco I and Sfu I and ligated into the plasmid pCRT7/VP22-1 (Invitrogen), which had been opened with restriction enzymes Nco I and Sfu I, and dephosphorylated with calf intestinal phosphatase. The resulting plasmid pCRT7-ΔVPCS (SEQ ID NO:15) was verified by DNA sequencing.

Expression of the fusion proteins in E. coli: E. coli BL21(DE3)-RIL cells (Stratagene) were transformed with pT7-TACS and grown on LB agar plates containing 100 µg/ml ampicillin. E. coli BL21(DE3)-RP cells (Stratagene) were transformed with pT7-VPCS and grown on LB agar plates containing 100 µg/ml ampicillin. E. coli BL21(DE3)-pLysS (Invitrogen) were transformed with pCRT7-ΔVPCS and grown on LB agar plates containing 25 µg/ml kanamycine and 34 µg/ml chloramphenicol. Single colonies were isolated and used to prepare glycerol stocks. Eight 5ml LB (Lura Bertani) aliquots containing antibiotics were inoculated with stabs from the glycerol stocks and grown overnight at 37°C with shaking. Two 5ml overnight cultures were each used to inoculate one of four 1L LB aliquots containing antibiotics and grown at 37°C with shaking. Growth rate was monitored by spectrophotometry at 578nm. When the cultures had obtained an $OD_{578} = 0,5$ expression of the fusion proteins were induced by the addition of 0,5 mM Isopropyl-β-D-1-thiogalactopyranosid (IPTG). Two hours after induction cells were harvested by centrifugation at 12000xg and the pellet rapidly frozen in liquid nitrogen and stored immediately at -80°C.

Purification of the fusion proteins from bacterial lysates: Each 10g cell pellet was resuspended on ice in 30ml Bicine buffer (50mM Bicine, pH 8,5) including one protease inhibitor tablet (Complete, Roche). Cells were lysed through threefold treatment (1500psi, 5 minutes) with the cell disruption bomb (Parr Instrument). 30ml of Benzonase (10000U, Merck) was added and cell extracts were incubated for 30 minutes at 4°C. Cell extracts were then centrifuged at 12,000xg (4°C). The pellet was redissolved in 8M urea, 50mM Bicine, 100mM DTT, pH 8,5 by incubation for 16 hours at 4°C. Protein extract was centrifuged at 31000xg and supernatant harvested. Protein extract was diluted in an equal volume of Chromatography buffer A (50mM Bicine, pH 8,5). PH was adjusted to pH

8,5 and the extract was filtered through a 0,45µm filter (Millipore). FPLC (Akta Explorer, Amersham Pharmacia) was performed using a cation exchange column (Sephacrose SP, Column body HR_5/5 (0.5 x 5cm), column volume (CV) 1ml, linear flow 300cm/hour, Amersham Pharmacia). After addition of sample to FPLC column, buffer was exchanged with Chromatography buffer A at 10 CV.

TAT-Cre and VP22-Cre fusion proteins were eluted from the column by gradient elution using chromatography buffer B (50mM Bicine, 1M NaCl, pH 8,5) using the following profile: 0 - 50 % buffer B, 0 CV; 50 % buffer B, 10 CV; 50 - 100 % buffer B (linear gradient), 20 CV; 100 % buffer B, 10 CV. ΔVP22-Cre protein was eluted from the column by gradient elution using the following profile: 0 - 10 % buffer B, 0 CV; 10 % buffer B, 10 CV; 10 - 30 % buffer B, 0 CV; 30 % buffer B, 10 CV; 30 - 100 % buffer B, 0 CV; 100 % buffer B, 10 CV. Three 1,5ml fractions each containing purified fusion proteins were collected. Purity and concentration of protein fractions were determined by Coomassie blue stained SDS-PAGE gels and Western blot analysis using dilutions of BSA standard solutions. In addition protein content was determined using a Bradford assay (Coomassie Plus protein assay, Pierce).

SDS-PAGE and Western blot analysis: SDS-PAGE and Coomassie staining was performed according to standard protocols (Maniatis et al., Cold Spring Harbor Laboratory, New York (1989)) using 4 - 12 % gradient SDS-polyacrylamide gels (NuPAGE, Invitrogen, cat. no.: NPO321). Western blot analysis was performed using a Semi-Try Blotting Chamber (Biorad) and nitrocellulose membranes (0,2 µm; Schleicher & Schuell) according to the manufacturers protocols. The fusion proteins were detected by using an alkaline phosphatase-conjugated anti-strep tag antibody (IBA, Cat. No.: 2-1503-001) according to the manufacturers protocol.

Generation of the M5Pax8 Cre reporter cell line: The SV40-transformed murine embryonic fibroblast line MEF5/5 (Schwenk et al., Nucl Acids Res 26(6), 1427-32 (1998)) was transfected with the vector pPGKpaX1 (Kellendonk et al, Nucl. Acids Res. 24, 1404-11 (1996)). 10^6 MEF5/5 cells were electroporated with 20 μ g pPGKpaX1 plasmid DNA linearised with Sca I and plated into 48-well-plates. The cells were cultured in DMEM/Glutamax medium (Life Technologies) supplemented with 10 % fetal calf serum at 37°C, 10 % CO₂ in humid atmosphere. Two days after transfection the medium was supplemented with 5 μ g/ml puromycine (Calbiochem) for the selection of stable integrants. 14 puromycine-resistant clones were expanded and tested by transient transfection with the Cre expression vector pPGK-Cre-pA (SEQ ID NO: 40). In two out of the 14 puromycine-resistant clones, the expression of β -galactosidase could be detected by staining with X-Gal. One of these clones, M5Pax8, was used as Cre reporter cell line.

Transfection and measurement of β -galactosidase activity: Fibroblasts (10^6 cells per 24 well plate (Falcon)) were transfected with 25 ng pCMV-I-Cre-pA (see SEQ ID NO:29) or pCMV-I- β -pA (see SEQ ID NO:30) plasmids using the FuGene transfection reagent (Roche Diagnostics). After 2 days the cells were lysed and the β -galactosidase activities were determined with the β -galactosidase reporter gene assay (Roche Diagnostics) according to the manufacturers guidelines using a Lumistar luminometer (MWG).

Histochemical detection of β -galactosidase activity: To quantitate β -galactosidase expression, fibroblast cells were washed once with phosphate buffered saline (PBS), and the cells were fixed for 5 minutes at room temperature in a solution of 4% formaldehyde in PBS. Next, the cells were washed twice with PBS and finally incubated in staining solution for 24 hours at 37°C (staining solution: 5 mM K₃(Fe(CN)₆), 5mM

K₄(Fe(CN)₆), 2mM MgCl₂, 1mg/ml X-Gal (BioMol) in PBS). Blue stained, β -galactosidase positive cells were detected and distinguished from negative (transparent) cells in a cell culture binocular microscope under 200x magnification. For each determination a minimum of 200 cells was counted.

PCR detection of Cre-mediated recombination: Genomic DNA extracted from tissue samples was subjected to PCR using Taq-polymerase (Gibco BRL Cat. No. 10342-020) using 20 pmol of each primer (sense: 5' -CAT CTC CGG GCC TTT CGA CCT G - 3', antisense: 5' -GCG ATC GGT GCG GGC CTC TTC - 3'; SEQ ID Nos: 41 and 42, respectively). PCR was performed using the following cycle profile: 2' 94°C, 35 x (30" 94°C, 30" 55 °C, 1' 72 °C), 10 min 72 °C. PCR products were separated on a 1,2 % agarose gel.

Example 1

The vector pT7-TACS (SEQ ID NO:16) was constructed for the expression of a TAT-Cre fusion protein in E. coli. The plasmid contains the coding region of the 11 amino acid protein transduction domain of the wild-type HIV TAT protein (Green and Loewenstein, Cell, 55(6):1179-88 (1988); Frankel and Pabo, Cell, 55(6): 1189-93 (1988); SEQ ID NO:10) fused to the N-terminus of Cre recombinase protein sequence. The 10-amino-acid strep tag at the C-terminus allows the detection and purification of the fusion protein using specific antibodies (Schmidt and Skerra, J. Chromatogr A 676: 337-345 (1994)). The protease factor Xa recognition site (Ile-Glu-Gly-Arg) permits the removal of the strep tag by proteolytic cleavage. The estimated molecular weight of the TAT-Cre fusion protein is 42 kDa. A scheme of the TAT-Cre expression vector is depicted in figure 2. For the expression of TAT-Cre, the E. coli strain BL21(DE3)-RIL (Stratagene) was used. This strain carries an IPTG-inducible T7 polymerase gene and additional copies of the tRNA genes for the 'rare

codons' argU, ileY and leuW.

E. coli BL21(DE3)-RIL cells were transformed with pT7-TACS and grown in LB medium containing 100 µg/ml ampicillin. The expression of the 40 kDa TAT-Cre fusion protein could be strongly induced by the addition of 0,5 mM IPTG to the culture medium. Analysis of protein lysates revealed that approximately 50 % of TAT-Cre protein accumulated as insoluble inclusion bodies. The inclusion bodies were extracted and dissolved in 8 M urea. TAT-Cre was subsequently purified from this fraction using ion exchange chromatography. The quantity and purity of TAT-Cre protein was determined using Coomassie stained SDS-PAGE gels and Western blot analysis (figure 3). The purification process yielded TAT-Cre protein extracts of 64 % purity and a concentration of 100 µg/ml.

To analyse the ability of the purified TAT-Cre protein to transduce into cultured cells, we used the fibroblast cell line M5Pax8 (R. Kühn, unpublished) that contains a loxP-containing reporter construct. This reporter, when recombined by Cre recombinase, allows the expression of a β -galactosidase gene (Buchholz et al, *Nucleic Acids Res.* 24, 4256-4262, 1996). Cells were cultured for 18 h with increasing concentrations of TAT-Cre protein in serum-free medium and analysed 4 days later for β -Galactosidase activity. Staining with X-Gal showed that > 50 % of the cells treated with 13,8 µg/ml TAT-Cre protein expressed β -galactosidase indicating recombination of the loxP-flanked reporter construct had occurred (figure 4). Measurement of β -galactosidase activity in cell lysates revealed an up to 30-fold higher level of β -galactosidase activity in comparison to cells which had been transiently transfected with an eukaryotic Cre expression vector (figure 5).

To investigate the activity of TAT-Cre protein in a living organism, we used a transgenic mouse strain carrying a loxP-flanked target for Cre-mediated recombination (Thorey et al., 1998, *Mol. Cell. Biol.* 18: 3081 – 3088). Mice were treated three times with intraperitoneal injections of 75 µg TAT Cre protein at two-day-intervals and analysed 2 days later. Genomic DNA was

isolated from a variety of organs and subjected to PCR amplification which specifically amplifies a 400 bp fragment of the recombined allele. The deleted allele could be detected in multiple tissues from treated mice indicating TAT-Cre-mediated recombination in these organs (figure 6). This experiments demonstrates that TAT-mediated delivery of active Cre protein works with sufficient efficacy to facilitate inducible gene targeting in cell lines and in living organisms.

Example 2

The vectors pT7-VPCS (SEQ ID NO:17) and pCRT7-ΔVPCS (SEQ ID NO:15) were constructed for the expression of VP22-Cre and ΔVP22-Cre fusion proteins in *E. coli*. The VP22-Cre gene of pT7-VPCS contains the full length protein translocation domain of the HSV VP22 protein (Elliott and O'Hare, Cell, 88(2): 223-33 (1987)), whereas the ΔVP22-Cre gene of pCRT7-ΔVPCS contains a truncated VP22 protein transduction domain (amino acids 159 – 301; Invitrogen; aa 16-157 of SEQ ID NO:14) fused to the N-terminus of Cre recombinase protein sequence. A 10-amino-acid strep tag at the C-terminus of Cre protein sequence allows the detection and purification of the fusion proteins using specific antibodies (Schmidt and Skerra, J. Chromatogr A 676: 337-345 (1994)). The protease factor Xa recognition site permits the removal of the Strep tag by proteolytic cleavage. The estimated molecular weight is 75 kDa for VP22-Cre protein and 60 kDa for ΔVP22-Cre protein. A scheme of the vectors pT7-VPCS and pCRT7-ΔVPCS is depicted in figure 7.

E. coli BL21(DE3)-RIP cells (Stratagene) were transformed with pT7-VPCS and cultured in LB medium containing 100 µg/ml ampicillin. *E. coli* BL21(DE3)-pLysS cells (Stratagene) were transformed with pCRT7-ΔVPCS and cultured in LB medium containing 25 µg/ml kanamycine and 34 µg/ml chloramphenicol. Expression of the VP22-Cre and ΔVP22-Cre fusion proteins could be induced by the addition of 0,5 mM IPTG to the culture medium. Analysis of protein extracts using Coomassie staining and

Western blotting of SDS-PAGE gels revealed that 50 - 60 % of VP22-Cre and Δ VP22-Cre proteins accumulated as insoluble inclusion bodies. The inclusion bodies were extracted and dissolved in 8 M urea. VP22-Cre and Δ VP22-Cre fusion proteins were subsequently purified using ion exchange chromatography. The quantity and purity of the isolated VP22-Cre and Δ VP22-Cre fusion proteins was determined using Coomassie stained SDS-PAGE gels and Western blot analysis (figure 8).

To analyse the ability of the purified fusion proteins to transduce into cultured cells, we used the fibroblast cell line M5Pax8 that contains a loxP-containing reporter construct. When recombined by Cre recombinase, the reporter allows the expression of a β -galactosidase gene (Buchholz et al, Nucleic Acids Res. 24, 4256-4262, 1996). The cells were cultured for 18 h with increasing concentrations of VP22-Cre and Δ VP22-Cre in serum-free medium and analysed 4 days later for β -Galactosidase activity. Staining with X-Gal showed \sim 2 % blue cells in the cultures treated with up to 15 μ g/ml Δ VP22-Cre indicating recombination of the loxP-flanked reporter construct had occurred. In contrast, cell cultures treated with up to 0,5 μ g/ml VP22-Cre did not show any X-gal staining (figure 9). Measurement of cell lysates revealed a strong increase of β -galactosidase activity upon Δ VP22-Cre treatment when compared to untreated cells (figure 10).

Genomic DNA was isolated and subjected to PCR amplification that specifically amplifies a 250 bp fragment of the recombined allele. The deleted allele could be detected in cells treated with both VP22-Cre and Δ VP22-Cre fusion proteins (figure 11).

This experiment demonstrates that VP22-mediated delivery of active Cre protein works with sufficient efficacy to facilitate inducible gene targeting.

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att aag aac gtt gat ccg aaa tca ttt aaa tta gtc caa aat aag tat Ile Lys Asn Val Asp Pro Lys Ser Phe Lys Leu Val Gln Asn Lys Tyr	672
210 215 220	
ctg gga gta ata atc cag tgt tta gtg aca gag aca aag aca agc gtt Leu Gly Val Ile Ile Gln Cys Leu Val Thr Glu Thr Lys Thr Ser Val	720
225 230 235 240	
agt agg cac ata tac ttc ttt agc gca agg ggt agg atc gat cca ctt Ser Arg His Ile Tyr Phe Phe Ser Ala Arg Gly Arg Ile Asp Pro Leu	768
245 250 255	
gta tat ttg gat gaa ttt ttg agg aat tct gaa cca gtc cta aaa cga Val Tyr Leu Asp Glu Phe Leu Arg Asn Ser Glu Pro Val Leu Lys Arg	816
260 265 270	
gta aat agg acc ggc aat tct tca agc aac aaa cag gaa tac caa tta Val Asn Arg Thr Gly Asn Ser Ser Ser Asn Lys Gln Glu Tyr Gln Leu	864
275 280 285	
tta aaa gat aac tta gtc aga tgc tac aac aag gct ttg aag aaa aat Leu Lys Asp Asn Leu Val Arg Ser Tyr Asn Lys Ala Leu Lys Lys Asn	912
290 295 300	
gcg cct tat cca atc ttt gct ata aag aat ggc cca aaa tct cac att Ala Pro Tyr Pro Ile Phe Ala Ile Lys Asn Gly Pro Lys Ser His Ile	960
305 310 315 320	
gga aga cat ttg atg acc tca ttt ctg tca atg aag ggc cta acg gag Gly Arg His Leu Met Thr Ser Phe Leu Ser Met Lys Gly Leu Thr Glu	1008
325 330 335	
ttg act aat gtt gtg gga aat tgg agc gat aag cgt gct tct gcc gtg Leu Thr Asn Val Val Gly Asn Trp Ser Asp Lys Arg Ala Ser Ala Val	1056
340 345 350	
gcc agg aca acg tat act cat cag ata aca gca ata cct gat cac tac Ala Arg Thr Thr Tyr Thr His Gln Ile Thr Ala Ile Pro Asp His Tyr	1104
355 360 365	
ttc gca cta gtt tct cgg tac tat gca tat gat cca ata tca aag gaa Phe Ala Leu Val Ser Arg Tyr Tyr Ala Tyr Asp Pro Ile Ser Lys Glu	1152
370 375 380	
atg ata gca ttg aag gat gag act aat cca att gag gag tgg cag cat Met Ile Ala Leu Lys Asp Glu Thr Asn Pro Ile Glu Glu Trp Gln His	1200
385 390 395 400	
ata gaa cag cta aag ggt agt gct gaa gga agc ata cga tac ccc gca Ile Glu Gln Leu Lys Gly Ser Ala Glu Gly Ser Ile Arg Tyr Pro Ala	1248
405 410 415	

tgg aat ggg ata ata tca cag gag gta cta gac tac ctt tca tcc tac 1296
 Trp Asn Gly Ile Ile Ser Gln Glu Val Leu Asp Tyr Leu Ser Ser Tyr
 420 425 430

ata aat aga cgc ata taatga 1317
 Ile Asn Arg Arg Ile
 435

<210> 4

<211> 437

<212> PRT

<213> Artificial Sequence

<223> Description of Artificial Sequence: DNA sequence
 coding for a fusion protein TAT-Flpe

<400> 4

Met Gly Tyr Gly Arg Lys Lys Arg Arg Gln Arg Arg Arg Gly Met Ser
 1 5 10 15

Gln Phe Asp Ile Leu Cys Lys Thr Pro Pro Lys Val Leu Val Arg Gln
 20 25 30

Phe Val Glu Arg Phe Glu Arg Pro Ser Gly Glu Lys Ile Ala Ser Cys
 35 40 45

Ala Ala Glu Leu Thr Tyr Leu Cys Trp Met Ile Thr His Asn Gly Thr
 50 55 60

Ala Ile Lys Arg Ala Thr Phe Met Ser Tyr Asn Thr Ile Ile Ser Asn
 65 70 75 80

Ser Leu Ser Phe Asp Ile Val Asn Lys Ser Leu Gln Phe Lys Tyr Lys
 85 90 95

Thr Gln Lys Ala Thr Ile Leu Glu Ala Ser Leu Lys Lys Leu Ile Pro
 100 105 110

Ala Trp Glu Phe Thr Ile Ile Pro Tyr Asn Gly Gln Lys His Gln Ser
 115 120 125

Asp Ile Thr Asp Ile Val Ser Ser Leu Gln Leu Gln Phe Glu Ser Ser
 130 135 140

Glu Glu Ala Asp Lys Gly Asn Ser His Ser Lys Lys Met Leu Lys Ala
 145 150 155 160

Leu Leu Ser Glu Gly Glu Ser Ile Trp Glu Ile Thr Glu Lys Ile Leu
 165 170 175

Asn Ser Phe Glu Tyr Thr Ser Arg Phe Thr Lys Thr Lys Thr Leu Tyr
 180 185 190

Gln Phe Leu Phe Leu Ala Thr Phe Ile Asn Cys Gly Arg Phe Ser Asp
 195 200 205

Ile Lys Asn Val Asp Pro Lys Ser Phe Lys Leu Val Gln Asn Lys Tyr
 210 215 220

Leu Gly Val Ile Ile Gln Cys Leu Val Thr Glu Thr Lys Thr Ser Val
 225 230 235 240

32

Ser Arg His Ile Tyr Phe Phe Ser Ala Arg Gly Arg Ile Asp Pro Leu
 245 250 255

Val Tyr Leu Asp Glu Phe Leu Arg Asn Ser Glu Pro Val Leu Lys Arg
 260 265 270

Val Asn Arg Thr Gly Asn Ser Ser Ser Asn Lys Gln Glu Tyr Gln Leu
 275 280 285

Leu Lys Asp Asn Leu Val Arg Ser Tyr Asn Lys Ala Leu Lys Lys Asn
 290 295 300

Ala Pro Tyr Pro Ile Phe Ala Ile Lys Asn Gly Pro Lys Ser His Ile
 305 310 315 320

Gly Arg His Leu Met Thr Ser Phe Leu Ser Met Lys Gly Leu Thr Glu
 325 330 335

Leu Thr Asn Val Val Gly Asn Trp Ser Asp Lys Arg Ala Ser Ala Val
 340 345 350

Ala Arg Thr Thr Tyr Thr His Gln Ile Thr Ala Ile Pro Asp His Tyr
 355 360 365

Phe Ala Leu Val Ser Arg Tyr Tyr Ala Tyr Asp Pro Ile Ser Lys Glu
 370 375 380

Met Ile Ala Leu Lys Asp Glu Thr Asn Pro Ile Glu Glu Trp Gln His
 385 390 395 400

Ile Glu Gln Leu Lys Gly Ser Ala Glu Gly Ser Ile Arg Tyr Pro Ala
 405 410 415

Trp Asn Gly Ile Ile Ser Gln Glu Val Leu Asp Tyr Leu Ser Ser Tyr
 420 425 430

Ile Asn Arg Arg Ile
 435

<210> 5

<211> 2004

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: DNA sequence
coding for a fusion protein VP22-Cre

<220>

<221> CDS

<222> (1)..(2001)

<400> 5

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Met Thr Ser Arg Arg Ser Val Lys Ser Gly Pro Arg Glu Val Pro Arg	
1 5 10 15	
gat gag tac gag gat ctg tac tac acc ccg tct tca ggt atg gcg agt	96
Asp Glu Tyr Glu Asp Leu Tyr Tyr Thr Pro Ser Ser Gly Met Ala Ser	
20 25 30	

ccc gat agt ccg cct gac acc tcc cgc cgt ggc gcc cta cag aca cgc Pro Asp Ser Pro Pro Asp Thr Ser Arg Arg Gly Ala Leu Gln Thr Arg	144
35 40 45	
tcg cgc cag agg ggc gag gtc cgt ttc gtc cag tac gac gag tcg gat Ser Arg Gln Arg Gly Glu Val Arg Phe Val Gln Tyr Asp Glu Ser Asp	192
50 55 60	
tat gcc ctc tac ggg ggc tcg tct tcc gaa gac gac gaa cac ccg gag Tyr Ala Leu Tyr Gly Gly Ser Ser Ser Glu Asp Asp Glu His Pro Glu	240
65 70 75 80	
gtc ccc cgg acg cgg cgt ccc gtt tcc ggg gcg gtt ttg tcc ggc ccg Val Pro Arg Thr Arg Arg Pro Val Ser Gly Ala Val Leu Ser Gly Pro	288
85 90 95	
ggg cct gcg cgg gcg cct ccg cca ccc gct ggg tcc gga ggg gcc gga Gly Pro Ala Arg Ala Pro Pro Pro Ala Gly Ser Gly Gly Ala Gly	336
100 105 110	
cgc aca ccc acc acc gcc ccc cgg gcc ccc cga acc cag cgg gtg gcg Arg Thr Pro Thr Thr Ala Pro Arg Ala Pro Arg Thr Gln Arg Val Ala	384
115 120 125	
act aag gcc ccc gcg gcc ccg gcg gcg gag acc acc cgc ggc agg aaa Thr Lys Ala Pro Ala Ala Pro Ala Ala Glu Thr Thr Arg Gly Arg Lys	432
130 135 140	
tcg gcc cag cca gaa tcc gcc gca ctc cca gac gcc ccc gcg tcg acg Ser Ala Gln Pro Glu Ser Ala Ala Leu Pro Asp Ala Pro Ala Ser Thr	480
145 150 155 160	
gcg cca acc cga tcc aag aca ccc gcg cag ggg ctg gcc aga aag ctg Ala Pro Thr Arg Ser Lys Thr Pro Ala Gln Gly Leu Ala Arg Lys Leu	528
165 170 175	
cac ttt agc acc gcc ccc cca aac ccc gac gcg cca tgg acc ccc cgg His Phe Ser Thr Ala Pro Pro Asn Pro Asp Ala Pro Trp Thr Pro Arg	576
180 185 190	
gtg gcc ggc ttt aac aag cgc gtc ttc tgc gcc gcg gtc ggg cgc ctg Val Ala Gly Phe Asn Lys Arg Val Phe Cys Ala Ala Val Gly Arg Leu	624
195 200 205	
gcg gcc atg cat gcc cgg atg gcg gcg gtc cag ctc tgg gac atg tcg Ala Ala Met His Ala Arg Met Ala Ala Val Gln Leu Trp Asp Met Ser	672
210 215 220	
cgt ccg cgc aca gac gaa gac ctc aac gaa ctc ctt ggc atc acc acc Arg Pro Arg Thr Asp Glu Asp Leu Asn Glu Leu Leu Gly Ile Thr Thr	720
225 230 235 240	
atc cgc gtg acg gtc tgc gag ggc aaa aac ctg ctt cag cgc gcc aac Ile Arg Val Thr Val Cys Glu Gly Lys Asn Leu Leu Gln Arg Ala Asn	768
245 250 255	
gag ttg gtg aat cca gac gtg gtg cag gac gtc gac gcg gcc acg gcg Glu Leu Val Asn Pro Asp Val Val Gln Asp Val Asp Ala Ala Thr Ala	816
260 265 270	
act cga ggg cgt tct gcg gcg tcg cgc ccc acc gag cga cct cga gcc Thr Arg Gly Arg Ser Ala Ala Ser Arg Pro Thr Glu Arg Pro Arg Ala	864
275 280 285	

cca gcc cgc tcc gct tct cgc ccc aga cgg ccc gtc gag ggt acc gag	912
Pro Ala Arg Ser Ala Ser Arg Pro Arg Arg Pro Val Glu Gly Thr Glu	
290 295 300	
ctc gga tcc act agt cca gtg tgg tgg aat tct gca gat atc cag cac	960
Leu Gly Ser Thr Ser Pro Val Trp Trp Asn Ser Ala Asp Ile Gln His	
305 310 315 320	
agt ggc ggc cgc atg tcc aat tta ctg acc gta cac caa aat ttg cct	1008
Ser Gly Gly Arg Met Ser Asn Leu Leu Thr Val His Gln Asn Leu Pro	
325 330 335	
gca tta ccg gtc gat gca acg agt gat gag gtt cgc aag aac ctg atg	1056
Ala Leu Pro Val Asp Ala Thr Ser Asp Glu Val Arg Lys Asn Leu Met	
340 345 350	
gac atg ttc agg gat cgc cag gcg ttt tct gag cat acc tgg aaa atg	1104
Asp Met Phe Arg Asp Arg Gln Ala Phe Ser Glu His Thr Trp Lys Met	
355 360 365	
ctt ctg tcc gtt tgc cgc tgc tgg gcg gca tgg tgc aag ttg aat aac	1152
Leu Leu Ser Val Cys Arg Ser Trp Ala Ala Trp Cys Lys Leu Asn Asn	
370 375 380	
cgg aaa tgg ttt ccc gca gaa cct gaa gat gtt cgc gat tat ctt cta	1200
Arg Lys Trp Phe Pro Ala Glu Pro Glu Asp Val Arg Asp Tyr Leu Leu	
385 390 395 400	
tat ctt cag gcg cgc ggt ctg gca gta aaa act atc cag caa cat ttg	1248
Tyr Leu Gln Ala Arg Gly Leu Ala Val Lys Thr Ile Gln Gln His Leu	
405 410 415	
ggc cag cta aac atg ctt cat cgt cgg tcc ggg ctg cca cga cca agt	1296
Gly Gln Leu Asn Met Leu His Arg Arg Ser Gly Leu Pro Arg Pro Ser	
420 425 430	
gac agc aat gct gtt tca ctg gtt atg cgg cgg atc cga aaa gaa aac	1344
Asp Ser Asn Ala Val Ser Leu Val Met Arg Arg Ile Arg Lys Glu Asn	
435 440 445	
gtt gat gcc ggt gaa cgt gca aaa cag gct cta gcg ttc gaa cgc act	1392
Val Asp Ala Gly Glu Arg Ala Lys Gln Ala Leu Ala Phe Glu Arg Thr	
450 455 460	
gat ttc gac cag gtt cgt tca ctc atg gaa aat agc gat cgc tgc cag	1440
Asp Phe Asp Gln Val Arg Ser Leu Met Glu Asn Ser Asp Arg Cys Gln	
465 470 475 480	
gat ata cgt aat ctg gca ttt ctg ggg att gct tat aac acc ctg tta	1488
Asp Ile Arg Asn Leu Ala Phe Leu Gly Ile Ala Tyr Asn Thr Leu Leu	
485 490 495	
cgt ata gcc gaa att gcc agg atc agg gtt aaa gat atc tca cgt act	1536
Arg Ile Ala Glu Ile Ala Arg Ile Arg Val Lys Asp Ile Ser Arg Thr	
500 505 510	
gac ggt ggg aga atg tta atc cat att ggc aga acg aaa acg ctg gtt	1584
Asp Gly Gly Arg Met Leu Ile His Ile Gly Arg Thr Lys Thr Leu Val	
515 520 525	
agc acc gca ggt gta gag aag gca ctt agc ctg ggg gta act aaa ctg	1632
Ser Thr Ala Gly Val Glu Lys Ala Leu Ser Leu Gly Val Thr Lys Leu	
530 535 540	

35

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gtc gag cga tgg att tcc gtc tct ggt gta gct gat gat ccg aat aac 1680
Val Glu Arg Trp Ile Ser Val Ser Gly Val Ala Asp Asp Pro Asn Asn
545                               550                               555                               560

tac ctg ttt tgc cgg gtc aga aaa aat ggt gtt gcc gcg cca tct gcc 1728
Tyr Leu Phe Cys Arg Val Arg Lys Asn Gly Val Ala Ala Pro Ser Ala
                    565                               570                               575

acc agc cag cta tca act cgc gcc ctg gaa ggg att ttt gaa gca act 1776
Thr Ser Gln Leu Ser Thr Arg Ala Leu Glu Gly Ile Phe Glu Ala Thr
                    580                               585                               590

cat cga ttg att tac ggc gct aag gat gac tct ggt cag aga tac ctg 1824
His Arg Leu Ile Tyr Gly Ala Lys Asp Asp Ser Gly Gln Arg Tyr Leu
                    595                               600                               605

gcc tgg tct gga cac agt gcc cgt gtc gga gcc gcg cga gat atg gcc 1872
Ala Trp Ser Gly His Ser Ala Arg Val Gly Ala Ala Arg Asp Met Ala
        610                               615                               620

cgc gct gga gtt tca ata ccg gag atc atg caa gct ggt ggc tgg acc 1920
Arg Ala Gly Val Ser Ile Pro Glu Ile Met Gln Ala Gly Gly Trp Thr
        625                               630                               635                               640

aat gta aat att gtc atg aac tat atc cgt aac ctg gat agt gaa aca 1968
Asn Val Asn Ile Val Met Asn Tyr Ile Arg Asn Leu Asp Ser Glu Thr
                    645                               650                               655

ggg gca atg gtg cgc ctg ctg gaa gat ggc gat tag 2004
Gly Ala Met Val Arg Leu Leu Glu Asp Gly Asp
                    660                               665

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<210> 6

<211> 667

<212> PRT

<213> Artificial Sequence

<223> Description of Artificial Sequence: DNA sequence
coding for a fusion protein VP22-Cre

<400> 6

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Asp Glu Tyr Glu Asp Leu Tyr Tyr Thr Pro Ser Ser Gly Met Ala Ser
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Pro Asp Ser Pro Pro Asp Thr Ser Arg Arg Gly Ala Leu Gln Thr Arg
                35                               40                               45

Ser Arg Gln Arg Gly Glu Val Arg Phe Val Gln Tyr Asp Glu Ser Asp
                50                               55                               60

Tyr Ala Leu Tyr Gly Gly Ser Ser Ser Glu Asp Asp Glu His Pro Glu
        65                               70                               75                               80

Val Pro Arg Thr Arg Arg Pro Val Ser Gly Ala Val Leu Ser Gly Pro
                85                               90                               95

Gly Pro Ala Arg Ala Pro Pro Pro Pro Ala Gly Ser Gly Gly Ala Gly
                100                               105                               110

Arg Thr Pro Thr Thr Ala Pro Arg Ala Pro Arg Thr Gln Arg Val Ala
                115                               120                               125

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Thr Lys Ala Pro Ala Ala Pro Ala Ala Glu Thr Thr Arg Gly Arg Lys
 130 135 140
 Ser Ala Gln Pro Glu Ser Ala Ala Leu Pro Asp Ala Pro Ala Ser Thr
 145 150 155 160
 Ala Pro Thr Arg Ser Lys Thr Pro Ala Gln Gly Leu Ala Arg Lys Leu
 165 170 175
 His Phe Ser Thr Ala Pro Pro Asn Pro Asp Ala Pro Trp Thr Pro Arg
 180 185 190
 Val Ala Gly Phe Asn Lys Arg Val Phe Cys Ala Ala Val Gly Arg Leu
 195 200 205
 Ala Ala Met His Ala Arg Met Ala Ala Val Gln Leu Trp Asp Met Ser
 210 215 220
 Arg Pro Arg Thr Asp Glu Asp Leu Asn Glu Leu Leu Gly Ile Thr Thr
 225 230 235 240
 Ile Arg Val Thr Val Cys Glu Gly Lys Asn Leu Leu Gln Arg Ala Asn
 245 250 255
 Glu Leu Val Asn Pro Asp Val Val Gln Asp Val Asp Ala Ala Thr Ala
 260 265 270
 Thr Arg Gly Arg Ser Ala Ala Ser Arg Pro Thr Glu Arg Pro Arg Ala
 275 280 285
 Pro Ala Arg Ser Ala Ser Arg Pro Arg Arg Pro Val Glu Gly Thr Glu
 290 295 300
 Leu Gly Ser Thr Ser Pro Val Trp Trp Asn Ser Ala Asp Ile Gln His
 305 310 315 320
 Ser Gly Gly Arg Met Ser Asn Leu Leu Thr Val His Gln Asn Leu Pro
 325 330 335
 Ala Leu Pro Val Asp Ala Thr Ser Asp Glu Val Arg Lys Asn Leu Met
 340 345 350
 Asp Met Phe Arg Asp Arg Gln Ala Phe Ser Glu His Thr Trp Lys Met
 355 360 365
 Leu Leu Ser Val Cys Arg Ser Trp Ala Ala Trp Cys Lys Leu Asn Asn
 370 375 380
 Arg Lys Trp Phe Pro Ala Glu Pro Glu Asp Val Arg Asp Tyr Leu Leu
 385 390 395 400
 Tyr Leu Gln Ala Arg Gly Leu Ala Val Lys Thr Ile Gln Gln His Leu
 405 410 415
 Gly Gln Leu Asn Met Leu His Arg Arg Ser Gly Leu Pro Arg Pro Ser
 420 425 430
 Asp Ser Asn Ala Val Ser Leu Val Met Arg Arg Ile Arg Lys Glu Asn
 435 440 445
 Val Asp Ala Gly Glu Arg Ala Lys Gln Ala Leu Ala Phe Glu Arg Thr
 450 455 460

37

Asp Phe Asp Gln Val Arg Ser Leu Met Glu Asn Ser Asp Arg Cys Gln
 465 470 475 480
 Asp Ile Arg Asn Leu Ala Phe Leu Gly Ile Ala Tyr Asn Thr Leu Leu
 485 490 495
 Arg Ile Ala Glu Ile Ala Arg Ile Arg Val Lys Asp Ile Ser Arg Thr
 500 505 510
 Asp Gly Gly Arg Met Leu Ile His Ile Gly Arg Thr Lys Thr Leu Val
 515 520 525
 Ser Thr Ala Gly Val Glu Lys Ala Leu Ser Leu Gly Val Thr Lys Leu
 530 535 540
 Val Glu Arg Trp Ile Ser Val Ser Gly Val Ala Asp Asp Pro Asn Asn
 545 550 555 560
 Tyr Leu Phe Cys Arg Val Arg Lys Asn Gly Val Ala Ala Pro Ser Ala
 565 570 575
 Thr Ser Gln Leu Ser Thr Arg Ala Leu Glu Gly Ile Phe Glu Ala Thr
 580 585 590
 His Arg Leu Ile Tyr Gly Ala Lys Asp Asp Ser Gly Gln Arg Tyr Leu
 595 600 605
 Ala Trp Ser Gly His Ser Ala Arg Val Gly Ala Ala Arg Asp Met Ala
 610 615 620
 Arg Ala Gly Val Ser Ile Pro Glu Ile Met Gln Ala Gly Gly Trp Thr
 625 630 635 640
 Asn Val Asn Ile Val Met Asn Tyr Ile Arg Asn Leu Asp Ser Glu Thr
 645 650 655
 Gly Ala Met Val Arg Leu Leu Glu Asp Gly Asp
 660 665

<210> 7

<211> 2247

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: DNA sequence
 coding for a fusion protein VP22-Flpe

<220>

<221> CDS

<222> (1)..(2241)

<400> 7

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Met Thr Ser Arg Arg Ser Val Lys Ser Gly Pro Arg Glu Val Pro Arg	
1 5 10 15	
gat gag tac gag gat ctg tac tac acc ccg tct tca ggt atg gcg agt	96
Asp Glu Tyr Glu Asp Leu Tyr Tyr Thr Pro Ser Ser Gly Met Ala Ser	
20 25 30	

ccc gat agt ccg cct gac acc tcc cgc cgt ggc gcc cta cag aca cgc	144
Pro Asp Ser Pro Pro Asp Thr Ser Arg Arg Gly Ala Leu Gln Thr Arg	
35 40 45	
tcg cgc cag agg ggc gag gtc cgt ttc gtc cag tac gac gag tcg gat	192
Ser Arg Gln Arg Gly Glu Val Arg Phe Val Gln Tyr Asp Glu Ser Asp	
50 55 60	
tat gcc ctc tac ggg ggc tcg tct tcc gaa gac gac gaa cac ccg gag	240
Tyr Ala Leu Tyr Gly Ser Ser Ser Glu Asp Asp Glu His Pro Glu	
65 70 75 80	
gtc ccc cgg acg ccg cgt ccc gtt tcc ggg gcg gtt ttg tcc ggc ccg	288
Val Pro Arg Thr Arg Arg Pro Val Ser Gly Ala Val Leu Ser Gly Pro	
85 90 95	
ggg cct gcg ccg gcg cct ccg cca ccc gct ggg tcc gga ggg gcc gga	336
Gly Pro Ala Arg Ala Pro Pro Pro Ala Gly Ser Gly Gly Ala Gly	
100 105 110	
cgc aca ccc acc acc gcc ccc ccg gcc ccc cga acc cag ccg gtg gcg	384
Arg Thr Pro Thr Thr Ala Pro Arg Ala Pro Arg Thr Gln Arg Val Ala	
115 120 125	
act aag gcc ccc gcg gcc ccg gcg gcg gag acc acc cgc gcc agg aaa	432
Thr Lys Ala Pro Ala Ala Pro Ala Ala Glu Thr Arg Gly Arg Lys	
130 135 140	
tcg gcc cag cca gaa tcc gcc gca ctc cca gac gcc ccc gcg tcg acg	480
Ser Ala Gln Pro Glu Ser Ala Ala Leu Pro Asp Ala Pro Ala Ser Thr	
145 150 155 160	
gcg cca acc cga tcc aag aca ccc gcg cag ggg ctg gcc aga aag ctg	528
Ala Pro Thr Arg Ser Lys Thr Pro Ala Gln Gly Leu Ala Arg Lys Leu	
165 170 175	
cac ttt agc acc gcc ccc cca aac ccc gac gcg cca tgg acc ccc ccg	576
His Phe Ser Thr Ala Pro Pro Asn Pro Asp Ala Pro Trp Thr Pro Arg	
180 185 190	
gtg gcc ggc ttt aac aag cgc gtc ttc tgc gcc gcg gtc ggg cgc ctg	624
Val Ala Gly Phe Asn Lys Arg Val Phe Cys Ala Ala Val Gly Arg Leu	
195 200 205	
gcg gcc atg cat gcc ccg atg gcg gcg gtc cag ctc tgg gac atg tcg	672
Ala Ala Met His Ala Arg Met Ala Ala Val Gln Leu Trp Asp Met Ser	
210 215 220	
cgt ccg cgc aca gac gaa gac ctc aac gaa ctc ctt ggc atc acc acc	720
Arg Pro Arg Thr Asp Glu Asp Leu Asn Glu Leu Leu Gly Ile Thr Thr	
225 230 235 240	
atc cgc gtg acg gtc tgc gag ggc aaa aac ctg ctt cag cgc gcc aac	768
Ile Arg Val Thr Val Cys Glu Gly Lys Asn Leu Leu Gln Arg Ala Asn	
245 250 255	
gag ttg gtg aat cca gac gtg gtg cag gac gtc gac gcg gcc acg gcg	816
Glu Leu Val Asn Pro Asp Val Val Gln Asp Val Asp Ala Ala Thr Ala	
260 265 270	
act cga ggg cgt tct gcg gcg tcg cgc ccc acc gag cga cct cga gcc	864
Thr Arg Gly Arg Ser Ala Ala Ser Arg Pro Thr Glu Arg Pro Arg Ala	
275 280 285	

cca gcc cgc tcc gct tct cgc ccc aga cgg ccc gtc gag ggt acc gag	912
Pro Ala Arg Ser Ala Ser Arg Pro Arg Arg Pro Val Glu Gly Thr Glu	
290 295 300	
ctc gga tcc act agt cca gtg tgg tgg aat tct gca gat atc cag cac	960
Leu Gly Ser Thr Ser Pro Val Trp Trp Asn Ser Ala Asp Ile Gln His	
305 310 315 320	
agt ggc ggc cgc atg agt caa ttt gat ata tta tgt aaa aca cca cct	1008
Ser Gly Gly Arg Met Ser Gln Phe Asp Ile Leu Cys Lys Thr Pro Pro	
325 330 335	
aag gtc ctg gtt cgt cag ttt gtg gaa agg ttt gaa aga cct tca ggg	1056
Lys Val Leu Val Arg Gln Phe Val Glu Arg Phe Glu Arg Pro Ser Gly	
340 345 350	
gaa aaa ata gca tca tgt gct gct gaa cta acc tat tta tgt tgg atg	1104
Glu Lys Ile Ala Ser Cys Ala Ala Glu Leu Thr Tyr Leu Cys Trp Met	
355 360 365	
att act cat aac gga aca gca atc aag aga gcc aca ttc atg agc tat	1152
Ile Thr His Asn Gly Thr Ala Ile Lys Arg Ala Thr Phe Met Ser Tyr	
370 375 380	
aat act atc ata agc aat tcg ctg agt ttc gat att gtc aac aaa tca	1200
Asn Thr Ile Ile Ser Asn Ser Leu Ser Phe Asp Ile Val Asn Lys Ser	
385 390 395 400	
ctc cag ttt aaa tac aag acg caa aaa gca aca att ctg gaa gcc tca	1248
Leu Gln Phe Lys Tyr Lys Thr Gln Lys Ala Thr Ile Leu Glu Ala Ser	
405 410 415	
tta aag aaa tta att cct gct tgg gaa ttt aca att att cct tac aat	1296
Leu Lys Lys Leu Ile Pro Ala Trp Glu Phe Thr Ile Ile Pro Tyr Asn	
420 425 430	
gga caa aaa cat caa tct gat atc act gat att gta agt agt ttg caa	1344
Gly Gln Lys His Gln Ser Asp Ile Thr Asp Ile Val Ser Ser Leu Gln	
435 440 445	
tta cag ttc gaa tca tcg gaa gaa gca gat aag gga aat agc cac agt	1392
Leu Gln Phe Glu Ser Ser Glu Glu Ala Asp Lys Gly Asn Ser His Ser	
450 455 460	
aaa aaa atg ctt aaa gca ctt cta agt gag ggt gaa agc atc tgg gag	1440
Lys Lys Met Leu Lys Ala Leu Leu Ser Glu Gly Glu Ser Ile Trp Glu	
465 470 475 480	
atc act gag aaa ata cta aat tcg ttt gag tat acc tcg aga ttt aca	1488
Ile Thr Glu Lys Ile Leu Asn Ser Phe Glu Tyr Thr Ser Arg Phe Thr	
485 490 495	
aaa aca aaa act tta tac caa ttc ctc ttc cta gct act ttc atc aat	1536
Lys Thr Lys Thr Leu Tyr Gln Phe Leu Phe Leu Ala Thr Phe Ile Asn	
500 505 510	
tgt gga aga ttc agc gat att aag aac gtt gat ccg aaa tca ttt aaa	1584
Cys Gly Arg Phe Ser Asp Ile Lys Asn Val Asp Pro Lys Ser Phe Lys	
515 520 525	
tta gtc caa aat aag tat ctg gga gta ata atc cag tgt tta gtg aca	1632
Leu Val Gln Asn Lys Tyr Leu Gly Val Ile Ile Gln Cys Leu Val Thr	
530 535 540	

40

gag aca aag aca agc gtt agt agg cac ata tac ttc ttt agc gca agg 1680
 Glu Thr Lys Thr Ser Val Ser Arg His Ile Tyr Phe Phe Ser Ala Arg
 545 550 555 560

ggt agg atc gat cca ctt gta tat ttg gat gaa ttt ttg agg aat tct 1728
 Gly Arg Ile Asp Pro Leu Val Tyr Leu Asp Glu Phe Leu Arg Asn Ser
 565 570 575

gaa cca gtc cta aaa cga gta aat agg acc ggc aat tct tca agc aac 1776
 Glu Pro Val Leu Lys Arg Val Asn Arg Thr Gly Asn Ser Ser Ser Asn
 580 585 590

aaa cag gaa tac caa tta tta aaa gat aac tta gtc aga tcg tac aac 1824
 Lys Gln Glu Tyr Gln Leu Leu Lys Asp Asn Leu Val Arg Ser Tyr Asn
 595 600 605

aag gct ttg aag aaa aat gcg cct tat cca atc ttt gct ata aag aat 1872
 Lys Ala Leu Lys Lys Asn Ala Pro Tyr Pro Ile Phe Ala Ile Lys Asn
 610 615 620

ggc cca aaa tct cac att gga aga cat ttg atg acc tca ttt ctg tca 1920
 Gly Pro Lys Ser His Ile Gly Arg His Leu Met Thr Ser Phe Leu Ser
 625 630 635 640

atg aag ggc cta acg gag ttg act aat gtt gtg gga aat tgg agc gat 1968
 Met Lys Gly Leu Thr Glu Leu Thr Asn Val Val Gly Asn Trp Ser Asp
 645 650 655

aag cgt gct tct gcc gtg gcc agg aca acg tat act cat cag ata aca 2016
 Lys Arg Ala Ser Ala Val Ala Arg Thr Thr Tyr Thr His Gln Ile Thr
 660 665 670

gca ata cct gat cac tac ttc gca cta gtt tct cgg tac tat gca tat 2064
 Ala Ile Pro Asp His Tyr Phe Ala Leu Val Ser Arg Tyr Tyr Ala Tyr
 675 680 685

gat cca ata tca aag gaa atg ata gca ttg aag gat gag act aat cca 2112
 Asp Pro Ile Ser Lys Glu Met Ile Ala Leu Lys Asp Glu Thr Asn Pro
 690 695 700

att gag gag tgg cag cat ata gaa cag cta aag ggt agt gct gaa gga 2160
 Ile Glu Glu Trp Gln His Ile Glu Gln Leu Lys Gly Ser Ala Glu Gly
 705 710 715 720

agc ata cga tac ccc gca tgg aat ggg ata ata tca cag gag gta cta 2208
 Ser Ile Arg Tyr Pro Ala Trp Asn Gly Ile Ile Ser Gln Glu Val Leu
 725 730 735

gac tac ctt tca tcc tac ata aat aga cgc ata taatga 2247
 Asp Tyr Leu Ser Ser Tyr Ile Asn Arg Arg Ile
 740 745

<210> 8

<211> 747

<212> PRT

<213> Artificial Sequence

 <223> Description of Artificial Sequence: DNA sequence
 coding for a fusion protein VP22-Flpe

<400> 8

 Met Thr Ser Arg Arg Ser Val Lys Ser Gly Pro Arg Glu Val Pro Arg
 1 5 10 15

41

Asp Glu Tyr Glu Asp Leu Tyr Tyr Thr Pro Ser Ser Gly Met Ala Ser
 20 25 30
 Pro Asp Ser Pro Pro Asp Thr Ser Arg Arg Gly Ala Leu Gln Thr Arg
 35 40 45
 Ser Arg Gln Arg Gly Glu Val Arg Phe Val Gln Tyr Asp Glu Ser Asp
 50 55 60
 Tyr Ala Leu Tyr Gly Gly Ser Ser Ser Glu Asp Asp Glu His Pro Glu
 65 70 75 80
 Val Pro Arg Thr Arg Arg Pro Val Ser Gly Ala Val Leu Ser Gly Pro
 85 90 95
 Gly Pro Ala Arg Ala Pro Pro Pro Pro Ala Gly Ser Gly Gly Ala Gly
 100 105 110
 Arg Thr Pro Thr Thr Ala Pro Arg Ala Pro Arg Thr Gln Arg Val Ala
 115 120 125
 Thr Lys Ala Pro Ala Ala Pro Ala Ala Glu Thr Thr Arg Gly Arg Lys
 130 135 140
 Ser Ala Gln Pro Glu Ser Ala Ala Leu Pro Asp Ala Pro Ala Ser Thr
 145 150 155 160
 Ala Pro Thr Arg Ser Lys Thr Pro Ala Gln Gly Leu Ala Arg Lys Leu
 165 170 175
 His Phe Ser Thr Ala Pro Pro Asn Pro Asp Ala Pro Trp Thr Pro Arg
 180 185 190
 Val Ala Gly Phe Asn Lys Arg Val Phe Cys Ala Ala Val Gly Arg Leu
 195 200 205
 Ala Ala Met His Ala Arg Met Ala Ala Val Gln Leu Trp Asp Met Ser
 210 215 220
 Arg Pro Arg Thr Asp Glu Asp Leu Asn Glu Leu Leu Gly Ile Thr Thr
 225 230 235 240
 Ile Arg Val Thr Val Cys Glu Gly Lys Asn Leu Leu Gln Arg Ala Asn
 245 250 255
 Glu Leu Val Asn Pro Asp Val Val Gln Asp Val Asp Ala Ala Thr Ala
 260 265 270
 Thr Arg Gly Arg Ser Ala Ala Ser Arg Pro Thr Glu Arg Pro Arg Ala
 275 280 285
 Pro Ala Arg Ser Ala Ser Arg Pro Arg Arg Pro Val Glu Gly Thr Glu
 290 295 300
 Leu Gly Ser Thr Ser Pro Val Trp Trp Asn Ser Ala Asp Ile Gln His
 305 310 315 320
 Ser Gly Gly Arg Met Ser Gln Phe Asp Ile Leu Cys Lys Thr Pro Pro
 325 330 335
 Lys Val Leu Val Arg Gln Phe Val Glu Arg Phe Glu Arg Pro Ser Gly
 340 345 350

42

Glu Lys Ile Ala Ser Cys Ala Ala Glu Leu Thr Tyr Leu Cys Trp Met
 355 360 365
 Ile Thr His Asn Gly Thr Ala Ile Lys Arg Ala Thr Phe Met Ser Tyr
 370 375 380
 Asn Thr Ile Ile Ser Asn Ser Leu Ser Phe Asp Ile Val Asn Lys Ser
 385 390 395 400
 Leu Gln Phe Lys Tyr Lys Thr Gln Lys Ala Thr Ile Leu Glu Ala Ser
 405 410 415
 Leu Lys Lys Leu Ile Pro Ala Trp Glu Phe Thr Ile Ile Pro Tyr Asn
 420 425 430
 Gly Gln Lys His Gln Ser Asp Ile Thr Asp Ile Val Ser Ser Leu Gln
 435 440 445
 Leu Gln Phe Glu Ser Ser Glu Glu Ala Asp Lys Gly Asn Ser His Ser
 450 455 460
 Lys Lys Met Leu Lys Ala Leu Leu Ser Glu Gly Glu Ser Ile Trp Glu
 465 470 475 480
 Ile Thr Glu Lys Ile Leu Asn Ser Phe Glu Tyr Thr Ser Arg Phe Thr
 485 490 495
 Lys Thr Lys Thr Leu Tyr Gln Phe Leu Phe Leu Ala Thr Phe Ile Asn
 500 505 510
 Cys Gly Arg Phe Ser Asp Ile Lys Asn Val Asp Pro Lys Ser Phe Lys
 515 520 525
 Leu Val Gln Asn Lys Tyr Leu Gly Val Ile Ile Gln Cys Leu Val Thr
 530 535 540
 Glu Thr Lys Thr Ser Val Ser Arg His Ile Tyr Phe Phe Ser Ala Arg
 545 550 555 560
 Gly Arg Ile Asp Pro Leu Val Tyr Leu Asp Glu Phe Leu Arg Asn Ser
 565 570 575
 Glu Pro Val Leu Lys Arg Val Asn Arg Thr Gly Asn Ser Ser Ser Asn
 580 585 590
 Lys Gln Glu Tyr Gln Leu Leu Lys Asp Asn Leu Val Arg Ser Tyr Asn
 595 600 605
 Lys Ala Leu Lys Lys Asn Ala Pro Tyr Pro Ile Phe Ala Ile Lys Asn
 610 615 620
 Gly Pro Lys Ser His Ile Gly Arg His Leu Met Thr Ser Phe Leu Ser
 625 630 635 640
 Met Lys Gly Leu Thr Glu Leu Thr Asn Val Val Gly Asn Trp Ser Asp
 645 650 655
 Lys Arg Ala Ser Ala Val Ala Arg Thr Thr Tyr Thr His Gln Ile Thr
 660 665 670
 Ala Ile Pro Asp His Tyr Phe Ala Leu Val Ser Arg Tyr Tyr Ala Tyr
 675 680 685

43

Asp Pro Ile Ser Lys Glu Met Ile Ala Leu Lys Asp Glu Thr Asn Pro
 690 695 700

Ile Glu Glu Trp Gln His Ile Glu Gln Leu Lys Gly Ser Ala Glu Gly
 705 710 715 720

Ser Ile Arg Tyr Pro Ala Trp Asn Gly Ile Ile Ser Gln Glu Val Leu
 725 730 735

Asp Tyr Leu Ser Ser Tyr Ile Asn Arg Arg Ile
 740 745

<210> 9
 <211> 33
 <212> DNA
 <213> Human immunodeficiency virus

<400> 9
 tacggccgca agaagcgccg ccaacgccgc cgc

33

<210> 10
 <211> 11
 <212> PRT
 <213> Human immunodeficiency virus

<400> 10
 Tyr Gly Arg Lys Lys Arg Arg Gln Arg Arg Arg
 1 5 10

<210> 11
 <211> 42
 <212> DNA
 <213> Human immunodeficiency virus

<220>
 <221> CDS
 <222> (4)..(42)

<400> 11
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 Gly Tyr Gly Arg Lys Lys Arg Arg Gln Arg Arg Arg Gly
 1 5 10

42

<210> 12
 <211> 13
 <212> PRT
 <213> Human immunodeficiency virus

<400> 12
 Gly Tyr Gly Arg Lys Lys Arg Arg Gln Arg Arg Arg Gly
 1 5 10

<210> 13
 <211> 1623
 <212> DNA
 <213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: DNA sequence
coding for a fusion protein deltaVP22cre-StrepTag

<220>

<221> CDS

<222> (1)..(1617)

<400> 13

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gcg cca acc cga tcc aag aca ccc gcg cag ggg ctg gcc aga aag ctg      96
Ala Pro Thr Arg Ser Lys Thr Pro Ala Gln Gly Leu Ala Arg Lys Leu
               20               25               30

cac ttt agc acc gcc ccc cca aac ccc gac gcg cca tgg acc ccc cgg     144
His Phe Ser Thr Ala Pro Pro Asn Pro Asp Ala Pro Trp Thr Pro Arg
               35               40               45

gtg gcc ggc ttt aac aag cgc gtc ttc tgc gcc gcg gtc ggg cgc ctg     192
Val Ala Gly Phe Asn Lys Arg Val Phe Cys Ala Ala Val Gly Arg Leu
   50               55               60

gcg gcc atg cat gcc cgg atg gcg gct gtc cag ctc tgg gac atg tcg     240
Ala Ala Met His Ala Arg Met Ala Ala Val Gln Leu Trp Asp Met Ser
   65               70               75               80

cgt ccg cgc aca gac gaa gac ctc aac gaa ctc ctt ggc atc acc acc     288
Arg Pro Arg Thr Asp Glu Asp Leu Asn Glu Leu Leu Gly Ile Thr Thr
               85               90               95

atc cgc gtg acg gtc tgc gag ggc aaa aac ctg ctt cag cgc gcc aac     336
Ile Arg Val Thr Val Cys Glu Gly Lys Asn Leu Leu Gln Arg Ala Asn
               100              105              110

gag ttg gtg aat cca gac gtg gtg cag gac gtc gac gcg gcc acg gcg     384
Glu Leu Val Asn Pro Asp Val Val Gln Asp Val Asp Ala Ala Thr Ala
               115              120              125

act cga ggg cgt tct gcg gcg tcg cgc ccc acc gag cga cct cga gcc     432
Thr Arg Gly Arg Ser Ala Ala Ser Arg Pro Thr Glu Arg Pro Arg Ala
               130              135              140

cca gcc cgc tcc gct tct cgc ccc aga cgg ccc gtc gag ggt acc gag     480
Pro Ala Arg Ser Ala Ser Arg Pro Arg Arg Pro Val Glu Gly Thr Glu
               145              150              155              160

ctc gga tcc act agt cca gtg tgg tgg aat tct gca gat atc cag cac     528
Leu Gly Ser Thr Ser Pro Val Trp Trp Asn Ser Ala Asp Ile Gln His
               165              170              175

agt ggc ggc cgc atg tcc aat tta ctg acc gta cac caa aat ttg cct     576
Ser Gly Gly Arg Met Ser Asn Leu Leu Thr Val His Gln Asn Leu Pro
               180              185              190

gca tta ccg gtc gat gca acg agt gat gag gtt cgc aag aac ctg atg     624
Ala Leu Pro Val Asp Ala Thr Ser Asp Glu Val Arg Lys Asn Leu Met
               195              200              205

gac atg ttc agg gat cgc cag gcg ttt tct gag cat acc tgg aaa atg     672
Asp Met Phe Arg Asp Arg Gln Ala Phe Ser Glu His Thr Trp Lys Met
               210              215              220

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45

ctt	ctg	tcc	gtt	tgc	cgg	tcg	tgg	gcg	gca	tgg	tgc	aag	ttg	aat	aac	720
Leu	Leu	Ser	Val	Cys	Arg	Ser	Trp	Ala	Ala	Trp	Cys	Lys	Leu	Asn	Asn	
225					230					235					240	
cgg	aaa	tgg	ttt	ccc	gca	gaa	cct	gaa	gat	gtt	cgc	gat	tat	ctt	cta	768
Arg	Lys	Trp	Phe	Pro	Ala	Glu	Pro	Glu	Asp	Val	Arg	Asp	Tyr	Leu	Leu	
				245					250					255		
tat	ctt	cag	gcg	cgc	ggt	ctg	gca	gta	aaa	act	atc	cag	caa	cat	ttg	816
Tyr	Leu	Gln	Ala	Arg	Gly	Leu	Ala	Val	Lys	Thr	Ile	Gln	Gln	His	Leu	
			260					265					270			
ggc	cag	cta	aac	atg	ctt	cat	cgt	cgg	tcc	ggg	ctg	cca	cga	cca	agt	864
Gly	Gln	Leu	Asn	Met	Leu	His	Arg	Arg	Ser	Gly	Leu	Pro	Arg	Pro	Ser	
		275					280					285				
gac	agc	aat	gct	gtt	tca	ctg	gtt	atg	cgg	cgg	atc	cga	aaa	gaa	aac	912
Asp	Ser	Asn	Ala	Val	Ser	Leu	Val	Met	Arg	Arg	Ile	Arg	Lys	Glu	Asn	
		290				295					300					
gtt	gat	gcc	ggt	gaa	cgt	gca	aaa	cag	gct	cta	gcg	ttc	gaa	cgc	act	960
Val	Asp	Ala	Gly	Glu	Arg	Ala	Lys	Gln	Ala	Leu	Ala	Phe	Glu	Arg	Thr	
305					310					315					320	
gat	ttc	gac	cag	gtt	cgt	tca	ctc	atg	gaa	aat	agc	gat	cgc	tgc	cag	1008
Asp	Phe	Asp	Gln	Val	Arg	Ser	Leu	Met	Glu	Asn	Ser	Asp	Arg	Cys	Gln	
				325					330					335		
gat	ata	cgt	aat	ctg	gca	ttt	ctg	ggg	att	gct	tat	aac	acc	ctg	tta	1056
Asp	Ile	Arg	Asn	Leu	Ala	Phe	Leu	Gly	Ile	Ala	Tyr	Asn	Thr	Leu	Leu	
			340					345					350			
cgt	ata	gcc	gaa	att	gcc	agg	atc	agg	gtt	aaa	gat	atc	tca	cgt	act	1104
Arg	Ile	Ala	Glu	Ile	Ala	Arg	Ile	Arg	Val	Lys	Asp	Ile	Ser	Arg	Thr	
		355					360					365				
gac	ggt	ggg	aga	atg	tta	atc	cat	att	ggc	aga	acg	aaa	acg	ctg	gtt	1152
Asp	Gly	Gly	Arg	Met	Leu	Ile	His	Ile	Gly	Arg	Thr	Lys	Thr	Leu	Val	
	370					375					380					
agc	acc	gca	ggt	gta	gag	aag	gca	ctt	agc	ctg	ggg	gta	act	aaa	ctg	1200
Ser	Thr	Ala	Gly	Val	Glu	Lys	Ala	Leu	Ser	Leu	Gly	Val	Thr	Lys	Leu	
					390					395					400	
gtc	gag	cga	tgg	att	tcc	gtc	tct	ggt	gta	gct	gat	gat	ccg	aat	aac	1248
Val	Glu	Arg	Trp	Ile	Ser	Val	Ser	Gly	Val	Ala	Asp	Asp	Pro	Asn	Asn	
				405					410					415		
tac	ctg	ttt	tgc	cgg	gtc	aga	aaa	aat	ggt	gtt	gcc	gcg	cca	tct	gcc	1296
Tyr	Leu	Phe	Cys	Arg	Val	Arg	Lys	Asn	Gly	Val	Ala	Ala	Pro	Ser	Ala	
			420					425					430			
acc	agc	cag	cta	tca	act	cgc	gcc	ctg	gaa	ggg	att	ttt	gaa	gca	act	1344
Thr	Ser	Gln	Leu	Ser	Thr	Arg	Ala	Leu	Glu	Gly	Ile	Phe	Glu	Ala	Thr	
		435					440					445				
cat	cga	ttg	att	tac	ggc	gct	aag	gat	gac	tct	ggt	cag	aga	tac	ctg	1392
His	Arg	Leu	Ile	Tyr	Gly	Ala	Lys	Asp	Asp	Ser	Gly	Gln	Arg	Tyr	Leu	
	450					455					460					
gcc	tgg	tct	gga	cac	agt	gcc	cgt	gtc	gga	gcc	gcg	cga	gat	atg	gcc	1440
Ala	Trp	Ser	Gly	His	Ser	Ala	Arg	Val	Gly	Ala	Ala	Arg	Asp	Met	Ala	
	465				470					475					480	

46

cgc gct gga gtt tca ata ccg gag atc atg caa gct ggt ggc tgg acc 1488
Arg Ala Gly Val Ser Ile Pro Glu Ile Met Gln Ala Gly Gly Trp Thr
485 490 495

aat gta aat att gtc atg aac tat atc cgt aac ctg gat agt gaa aca 1536
Asn Val Asn Ile Val Met Asn Tyr Ile Arg Asn Leu Asp Ser Glu Thr
500 505 510

ggg gca atg gtg cgc ctg ctg gaa gat ggc gat ggt atc gaa ggt cgt 1584
Gly Ala Met Val Arg Leu Leu Glu Asp Gly Asp Gly Ile Glu Gly Arg
515 520 525

ggt agc gct tgg cgt cac ccg cag ttc ggt ggt taataa 1623
Gly Ser Ala Trp Arg His Pro Gln Phe Gly Gly
530 535

<210> 14

<211> 539

<212> PRT

<213> Artificial Sequence

<223> Description of Artificial Sequence: DNA sequence coding for a fusion protein deltaVP22cre-StrepTag

<400> 14

Met Ala Ser Met Thr Gly Gly Gln Gln Met Gly Arg Asp Pro Ser Thr
1 5 10 15

Ala Pro Thr Arg Ser Lys Thr Pro Ala Gln Gly Leu Ala Arg Lys Leu
20 25 30

His Phe Ser Thr Ala Pro Pro Asn Pro Asp Ala Pro Trp Thr Pro Arg
35 40 45

Val Ala Gly Phe Asn Lys Arg Val Phe Cys Ala Ala Val Gly Arg Leu
50 55 60

Ala Ala Met His Ala Arg Met Ala Ala Val Gln Leu Trp Asp Met Ser
65 70 75 80

Arg Pro Arg Thr Asp Glu Asp Leu Asn Glu Leu Leu Gly Ile Thr Thr
85 90 95

Ile Arg Val Thr Val Cys Glu Gly Lys Asn Leu Leu Gln Arg Ala Asn
100 105 110

Glu Leu Val Asn Pro Asp Val Val Gln Asp Val Asp Ala Ala Thr Ala
115 120 125

Thr Arg Gly Arg Ser Ala Ala Ser Arg Pro Thr Glu Arg Pro Arg Ala
130 135 140

Pro Ala Arg Ser Ala Ser Arg Pro Arg Arg Pro Val Glu Gly Thr Glu
145 150 155 160

Leu Gly Ser Thr Ser Pro Val Trp Trp Asn Ser Ala Asp Ile Gln His
165 170 175

Ser Gly Gly Arg Met Ser Asn Leu Leu Thr Val His Gln Asn Leu Pro
180 185 190

Ala Leu Pro Val Asp Ala Thr Ser Asp Glu Val Arg Lys Asn Leu Met
195 200 205

47

Asp Met Phe Arg Asp Arg Gln Ala Phe Ser Glu His Thr Trp Lys Met
 210 215 220
 Leu Leu Ser Val Cys Arg Ser Trp Ala Ala Trp Cys Lys Leu Asn Asn
 225 230 235 240
 Arg Lys Trp Phe Pro Ala Glu Pro Glu Asp Val Arg Asp Tyr Leu Leu
 245 250 255
 Tyr Leu Gln Ala Arg Gly Leu Ala Val Lys Thr Ile Gln Gln His Leu
 260 265 270
 Gly Gln Leu Asn Met Leu His Arg Arg Ser Gly Leu Pro Arg Pro Ser
 275 280 285
 Asp Ser Asn Ala Val Ser Leu Val Met Arg Arg Ile Arg Lys Glu Asn
 290 295 300
 Val Asp Ala Gly Glu Arg Ala Lys Gln Ala Leu Ala Phe Glu Arg Thr
 305 310 315 320
 Asp Phe Asp Gln Val Arg Ser Leu Met Glu Asn Ser Asp Arg Cys Gln
 325 330 335
 Asp Ile Arg Asn Leu Ala Phe Leu Gly Ile Ala Tyr Asn Thr Leu Leu
 340 345 350
 Arg Ile Ala Glu Ile Ala Arg Ile Arg Val Lys Asp Ile Ser Arg Thr
 355 360 365
 Asp Gly Gly Arg Met Leu Ile His Ile Gly Arg Thr Lys Thr Leu Val
 370 375 380
 Ser Thr Ala Gly Val Glu Lys Ala Leu Ser Leu Gly Val Thr Lys Leu
 385 390 395 400
 Val Glu Arg Trp Ile Ser Val Ser Gly Val Ala Asp Asp Pro Asn Asn
 405 410 415
 Tyr Leu Phe Cys Arg Val Arg Lys Asn Gly Val Ala Ala Pro Ser Ala
 420 425 430
 Thr Ser Gln Leu Ser Thr Arg Ala Leu Glu Gly Ile Phe Glu Ala Thr
 435 440 445
 His Arg Leu Ile Tyr Gly Ala Lys Asp Asp Ser Gly Gln Arg Tyr Leu
 450 455 460
 Ala Trp Ser Gly His Ser Ala Arg Val Gly Ala Ala Arg Asp Met Ala
 465 470 475 480
 Arg Ala Gly Val Ser Ile Pro Glu Ile Met Gln Ala Gly Gly Trp Thr
 485 490 495
 Asn Val Asn Ile Val Met Asn Tyr Ile Arg Asn Leu Asp Ser Glu Thr
 500 505 510
 Gly Ala Met Val Arg Leu Leu Glu Asp Gly Asp Gly Ile Glu Gly Arg
 515 520 525
 Gly Ser Ala Trp Arg His Pro Gln Phe Gly Gly
 530 535

<210> 15
 <211> 5953
 <212> DNA
 <213> Artificial Sequence

<220>
 <223> Description of Artificial Sequence: vector
 pCRT7-deltaVPCS

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 gggattttttg aagcaactca tcgattgatt tacggcgcta aggatgactc tggtcagaga 180
 tacctggcct ggtctggaca cagtgcccggt gtccggagccg cgcgagatat ggcccgcgct 240
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 aactatatcc gtaacctgga tagtgaaaca ggggcaatgg tgcgcctgct ggaagatggc 360
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 tcgaacaaaa actcatctca gaagaggatc tgaatatgca tacgggtcat catcaccatc 480
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 taagggggat ttctgttcat gggggtaatg ataccgatga aacgagagag gatgctcacg 3180
 atacgggtta ctgatgatga acatgcccgg ttactggaac gttgtgaggg taaacaactg 3240

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<210> 16

<211> 4727

<212> DNA

<213> Artificial Sequence

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<223> Description of Artificial Sequence: vector
pT7-TACS

<400> 16

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<210> 17

<211> 4488

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: vector
pT7-VPCS

<400> 17

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<211> 1125

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: DNA sequence
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<220>

<221> CDS

<222> (1)..(1119)

<400> 18

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Asn Leu Leu Thr Val His Gln Asn Leu Pro Ala Leu Pro Val Asp Ala
20 25 30

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Thr Ser Asp Glu Val Arg Lys Asn Leu Met Asp Met Phe Arg Asp Arg
35 40 45

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cag gcg ttt tct gag cat acc tgg aaa atg ctt ctg tcc gtt tgc cgg 192
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50 55 60

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53

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ttt Phe	ctg Leu	ggg Gly	att Ile 180	gct Ala	tat Tyr	aac Asn	acc Thr	ctg Leu 185	tta Leu	cgt Arg	ata Ile	gcc Ala	gaa Glu 190	att Ile	gcc Ala	576
agg Arg	atc Ile	agg Arg 195	gtt Val	aaa Lys	gat Asp	atc Ile	tca Ser 200	cgt Arg	act Thr	gac Asp	ggt Gly	ggg Gly 205	aga Arg	atg Met	tta Leu	624
atc Ile	cat His 210	att Ile	ggc Gly	aga Arg	acg Thr	aaa Lys 215	acg Thr	ctg Leu	gtt Val	agc Ser	acc Thr 220	gca Ala	ggt Gly	gta Val	gag Glu	672
aag Lys 225	gca Ala	ctt Leu	agc Ser	ctg Leu	ggg Gly 230	gta Val	act Thr	aaa Lys	ctg Leu	gtc Val 235	gag Glu	cga Arg	tg Trp	att Ile	tcc Ser 240	720
gtc Val	tct Ser	ggt Gly	gta Val	gct Ala 245	gat Asp	gat Asp	ccg Pro	aat Asn	aac Asn 250	tac Tyr	ctg Leu	ttt Phe	tgc Cys	cgg Arg 255	gtc Val	768
aga Arg	aaa Lys	aat Asn	ggt Gly 260	gtt Val	gcc Ala	gcg Ala	cca Pro	tct Ser 265	gcc Ala	acc Thr	agc Ser	cag Gln	cta Leu 270	tca Ser	act Thr	816
cgc Arg	gcc Ala	ctg Leu 275	gaa Glu	ggg Gly	att Ile	ttt Phe	gaa Glu 280	gca Ala	act Thr	cat His	cga Arg	ttg Leu 285	att Ile	tac Tyr	ggc Gly	864
gct Ala	aag Lys 290	gat Asp	gac Asp	tct Ser	ggt Gly	cag Gln 295	aga Arg	tac Tyr	ctg Leu	gcc Ala	tg Trp 300	tct Ser	gga Gly	cac His	agt Ser	912
gcc Ala 305	cgt Arg	gtc Val	gga Gly	gcc Ala	gcg Ala 310	cga Arg	gat Asp	atg Met	gcc Ala	cgc Arg 315	gct Ala	gga Gly	gtt Val	tca Ser	ata Ile 320	960

54

ccg gag atc atg caa gct ggt ggc tgg acc aat gta aat att gtc atg 1008
 Pro Glu Ile Met Gln Ala Gly Gly Trp Thr Asn Val Asn Ile Val Met
 325 330 335
 aac tat atc cgt aac ctg gat agt gaa aca ggg gca atg gtg cgc ctg 1056
 Asn Tyr Ile Arg Asn Leu Asp Ser Glu Thr Gly Ala Met Val Arg Leu
 340 345 350
 ctg gaa gat ggc gat ggt atc gaa ggt cgt ggt agc gct tgg cgt cac 1104
 Leu Glu Asp Gly Asp Gly Ile Glu Gly Arg Gly Ser Ala Trp Arg His
 355 360 365
 ccg cag ttc ggt ggt taataa 1125
 Pro Gln Phe Gly Gly
 370

<210> 19

<211> 373

<212> PRT

<213> Artificial Sequence

<223> Description of Artificial Sequence: DNA sequence
 coding for a fusion protein TATcreStrepTag

<400> 19

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 Asn Leu Leu Thr Val His Gln Asn Leu Pro Ala Leu Pro Val Asp Ala
 20 25 30
 Thr Ser Asp Glu Val Arg Lys Asn Leu Met Asp Met Phe Arg Asp Arg
 35 40 45
 Gln Ala Phe Ser Glu His Thr Trp Lys Met Leu Leu Ser Val Cys Arg
 50 55 60
 Ser Trp Ala Ala Trp Cys Lys Leu Asn Asn Arg Lys Trp Phe Pro Ala
 65 70 75 80
 Glu Pro Glu Asp Val Arg Asp Tyr Leu Leu Tyr Leu Gln Ala Arg Gly
 85 90 95
 Leu Thr Val Lys Thr Ile Gln Gln His Leu Gly Gln Leu Asn Met Leu
 100 105 110
 His Arg Arg Ser Gly Leu Pro Arg Pro Ser Asp Ser Asn Ala Val Ser
 115 120 125
 Leu Val Met Arg Arg Ile Arg Lys Glu Asn Val Asp Ala Gly Glu Arg
 130 135 140
 Ala Lys Gln Ala Leu Ala Phe Glu Arg Thr Asp Phe Asp Gln Val Arg
 145 150 155 160
 Ser Leu Met Glu Asn Ser Asp Arg Cys Gln Asp Ile Arg Asn Leu Ala
 165 170 175
 Phe Leu Gly Ile Ala Tyr Asn Thr Leu Leu Arg Ile Ala Glu Ile Ala
 180 185 190
 Arg Ile Arg Val Lys Asp Ile Ser Arg Thr Asp Gly Gly Arg Met Leu
 195 200 205

55

Ile His Ile Gly Arg Thr Lys Thr Leu Val Ser Thr Ala Gly Val Glu
 210 215 220

Lys Ala Leu Ser Leu Gly Val Thr Lys Leu Val Glu Arg Trp Ile Ser
 225 230 235 240

Val Ser Gly Val Ala Asp Asp Pro Asn Asn Tyr Leu Phe Cys Arg Val
 245 250 255

Arg Lys Asn Gly Val Ala Ala Pro Ser Ala Thr Ser Gln Leu Ser Thr
 260 265 270

Arg Ala Leu Glu Gly Ile Phe Glu Ala Thr His Arg Leu Ile Tyr Gly
 275 280 285

Ala Lys Asp Asp Ser Gly Gln Arg Tyr Leu Ala Trp Ser Gly His Ser
 290 295 300

Ala Arg Val Gly Ala Ala Arg Asp Met Ala Arg Ala Gly Val Ser Ile
 305 310 315 320

Pro Glu Ile Met Gln Ala Gly Gly Trp Thr Asn Val Asn Ile Val Met
 325 330 335

Asn Tyr Ile Arg Asn Leu Asp Ser Glu Thr Gly Ala Met Val Arg Leu
 340 345 350

Leu Glu Asp Gly Asp Gly Ile Glu Gly Arg Gly Ser Ala Trp Arg His
 355 360 365

Pro Gln Phe Gly Gly
 370

<210> 20

<211> 2055

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: DNA sequence
coding for a fusion protein VP22creStrepTag

<220>

<221> CDS

<222> (1)..(2049)

<400> 20

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1 5 10 15	
gat gag tac gag gat ctg tac tac acc ccg tct tca ggt atg gcg agt	96
Asp Glu Tyr Glu Asp Leu Tyr Tyr Thr Pro Ser Ser Gly Met Ala Ser	
20 25 30	
ccc gat agt ccg cct gac acc tcc cgc cgt ggc gcc cta cag aca cgc	144
Pro Asp Ser Pro Pro Asp Thr Ser Arg Arg Gly Ala Leu Gln Thr Arg	
35 40 45	
tcg cgc cag agg ggc gag gtc cgt ttc gtc cag tac gac gag tcg gat	192
Ser Arg Gln Arg Gly Glu Val Arg Phe Val Gln Tyr Asp Glu Ser Asp	
50 55 60	

56

tat gcc ctc tac ggg ggc tgc tct tcc gaa gac gac gaa cac ccg gag	240
Tyr Ala Leu Tyr Gly Gly Ser Ser Ser Glu Asp Asp Glu His Pro Glu	
65 70 75 80	
gtc ccc cgg acg cgg cgt ccc gtt tcc ggg gcg gtt ttg tcc ggc ccg	288
Val Pro Arg Thr Arg Arg Pro Val Ser Gly Ala Val Leu Ser Gly Pro	
85 90 95	
ggg cct gcg cgg gcg cct ccg cca ccc gct ggg tcc gga ggg gcc gga	336
Gly Pro Ala Arg Ala Pro Pro Pro Ala Gly Ser Gly Ala Ala Gly	
100 105 110	
cgc aca ccc acc acc gcc ccc cgg gcc ccc cga acc cag cgg gtg gcg	384
Arg Thr Pro Thr Thr Ala Pro Arg Ala Pro Arg Thr Gln Arg Val Ala	
115 120 125	
tct aag gcc ccc gcg gcc ccg gcg gcg gag acc acc cgc ggc agg aaa	432
Ser Lys Ala Pro Ala Ala Pro Ala Ala Glu Thr Thr Arg Gly Arg Lys	
130 135 140	
tgc gcc cag cca gaa tcc gcc gca ctc cca gac gcc ccc gcg tgc acg	480
Ser Ala Gln Pro Glu Ser Ala Ala Leu Pro Asp Ala Pro Ala Ser Thr	
145 150 155 160	
gcg cca acc cga tcc aag aca ccc gcg cag ggg ctg gcc aga aag ctg	528
Ala Pro Thr Arg Ser Lys Thr Pro Ala Gln Gly Leu Ala Arg Lys Leu	
165 170 175	
cac ttt agc acc gcc ccc cca aac ccc gac gcg cca tgg acc ccc cgg	576
His Phe Ser Thr Ala Pro Pro Asn Pro Asp Ala Pro Trp Thr Pro Arg	
180 185 190	
gtg gcc ggc ttt aac aag cgc gtc ttc tgc gcc gcg gtc ggg cgc ctg	624
Val Ala Gly Phe Asn Lys Arg Val Phe Cys Ala Ala Val Gly Arg Leu	
195 200 205	
gcg gcc atg cat gcc cgg atg gcg gct gtc cag ctc tgg gac atg tgc	672
Ala Ala Met His Ala Arg Met Ala Val Gln Leu Trp Asp Met Ser	
210 215 220	
cgt ccg cgc aca gac gaa gac ctc aac gaa ctc ctt ggc atc acc acc	720
Arg Pro Arg Thr Asp Glu Asp Leu Asn Glu Leu Leu Gly Ile Thr Thr	
225 230 235 240	
atc cgc gtg acg gtc tgc gag ggc aaa aac ctg ctt cag cgc gcc aac	768
Ile Arg Val Thr Val Cys Glu Gly Lys Asn Leu Leu Gln Arg Ala Asn	
245 250 255	
gag ttg gtg aat cca gac gtg gtg cag gac gtc gac gcg gcc acg gcg	816
Glu Leu Val Asn Pro Asp Val Val Gln Asp Val Asp Ala Ala Thr Ala	
260 265 270	
act cga ggg cgt tct gcg gcg tgc cgc ccc acc gag cga cct cga gcc	864
Thr Arg Gly Arg Ser Ala Ala Ser Arg Pro Thr Glu Arg Pro Arg Ala	
275 280 285	
cca gcc cgc tcc gct tct cgc ccc aga cgg ccc gtc gag ggt acc gag	912
Pro Ala Arg Ser Ala Ser Arg Pro Arg Arg Pro Val Glu Gly Thr Glu	
290 295 300	
ctc gga tcc act agt cca gtg tgg tgg aat tct gca gat atc cag cac	960
Leu Gly Ser Thr Ser Pro Val Trp Trp Asn Ser Ala Asp Ile Gln His	
305 310 315 320	

57

agt ggc ggc cgc atg tcc aat tta ctg acc gta cac caa aat ttg cct	1008
Ser Gly Gly Arg Met Ser Asn Leu Leu Thr Val His Gln Asn Leu Pro	
325 330 335	
gca tta ccg gtc gat gca acg agt gat gag gtt cgc aag aac ctg atg	1056
Ala Leu Pro Val Asp Ala Thr Ser Asp Glu Val Arg Lys Asn Leu Met	
340 345 350	
gac atg ttc agg gat cgc cag gcg ttt tct gag cat acc tgg aaa atg	1104
Asp Met Phe Arg Asp Arg Gln Ala Phe Ser Glu His Thr Trp Lys Met	
355 360 365	
ctt ctg tcc gtt tgc cgg tgc tgg gcg gca tgg tgc aag ttg aat aac	1152
Leu Leu Ser Val Cys Arg Ser Trp Ala Ala Trp Cys Lys Leu Asn Asn	
370 375 380	
cgg aaa tgg ttt ccc gca gaa cct gaa gat gtt cgc gat tat ctt cta	1200
Arg Lys Trp Phe Pro Ala Glu Pro Glu Asp Val Arg Asp Tyr Leu Leu	
385 390 395 400	
tat ctt cag gcg cgc ggt ctg gca gta aaa act atc cag caa cat ttg	1248
Tyr Leu Gln Ala Arg Gly Leu Ala Val Lys Thr Ile Gln Gln His Leu	
405 410 415	
ggc cag cta aac atg ctt cat cgt cgg tcc ggg ctg cca cga cca agt	1296
Gly Gln Leu Asn Met Leu His Arg Arg Ser Gly Leu Pro Arg Pro Ser	
420 425 430	
gac agc aat gct gtt tca ctg gtt atg cgg cgg atc cga aaa gaa aac	1344
Asp Ser Asn Ala Val Ser Leu Val Met Arg Arg Ile Arg Lys Glu Asn	
435 440 445	
gtt gat gcc ggt gaa cgt gca aaa cag gct cta gcg ttc gaa cgc act	1392
Val Asp Ala Gly Glu Arg Ala Lys Gln Ala Leu Ala Phe Glu Arg Thr	
450 455 460	
gat ttc gac cag gtt cgt tca ctc atg gaa aat agc gat cgc tgc cag	1440
Asp Phe Asp Gln Val Arg Ser Leu Met Glu Asn Ser Asp Arg Cys Gln	
465 470 475 480	
gat ata cgt aat ctg gca ttt ctg ggg att gct tat aac acc ctg tta	1488
Asp Ile Arg Asn Leu Ala Phe Leu Gly Ile Ala Tyr Asn Thr Leu Leu	
485 490 495	
cgt ata gcc gaa att gcc agg atc agg gtt aaa gat atc tca cgt act	1536
Arg Ile Ala Glu Ile Ala Arg Ile Arg Val Lys Asp Ile Ser Arg Thr	
500 505 510	
gac ggt ggg aga atg tta atc cat att ggc aga acg aaa acg ctg gtt	1584
Asp Gly Gly Arg Met Leu Ile His Ile Gly Arg Thr Lys Thr Leu Val	
515 520 525	
agc acc gca ggt gta gag aag gca ctt agc ctg ggg gta act aaa ctg	1632
Ser Thr Ala Gly Val Glu Lys Ala Leu Ser Leu Gly Val Thr Lys Leu	
530 535 540	
gtc gag cga tgg att tcc gtc tct ggt gta gct gat gat ccg aat aac	1680
Val Glu Arg Trp Ile Ser Val Ser Gly Val Ala Asp Asp Pro Asn Asn	
545 550 555 560	
tac ctg ttt tgc cgg gtc aga aaa aat ggt gtt gcc gcg cca tct gcc	1728
Tyr Leu Phe Cys Arg Val Arg Lys Asn Gly Val Ala Ala Pro Ser Ala	
565 570 575	

58

acc agc cag cta tca act cgc gcc ctg gaa ggg att ttt gaa gca act 1776
 Thr Ser Gln Leu Ser Thr Arg Ala Leu Glu Gly Ile Phe Glu Ala Thr
 580 585 590

cat cga ttg att tac ggc gct aag gat gac tct ggt cag aga tac ctg 1824
 His Arg Leu Ile Tyr Gly Ala Lys Asp Asp Ser Gly Gln Arg Tyr Leu
 595 600 605

gcc tgg tct gga cac agt gcc cgt gtc gga gcc gcg cga gat atg gcc 1872
 Ala Trp Ser Gly His Ser Ala Arg Val Gly Ala Ala Arg Asp Met Ala
 610 615 620

cgc gct gga gtt tca ata ccg gag atc atg caa gct ggt ggc tgg acc 1920
 Arg Ala Gly Val Ser Ile Pro Glu Ile Met Gln Ala Gly Gly Trp Thr
 625 630 635 640

aat gta aat att gtc atg aac tat atc cgt aac ctg gat agt gaa aca 1968
 Asn Val Asn Ile Val Met Asn Tyr Ile Arg Asn Leu Asp Ser Glu Thr
 645 650 655

ggg gca atg gtg cgc ctg ctg gaa gat ggc gat ggt atc gaa ggt cgt 2016
 Gly Ala Met Val Arg Leu Leu Glu Asp Gly Asp Gly Ile Glu Gly Arg
 660 665 670

ggt agc gct tgg cgt cac ccg cag ttc ggt ggt taataa 2055
 Gly Ser Ala Trp Arg His Pro Gln Phe Gly Gly
 675 680

<210> 21

<211> 683

<212> PRT

<213> Artificial Sequence

<223> Description of Artificial Sequence: DNA sequence
 coding for a fusion protein VP22creStrepTag

<400> 21

Met Thr Ser Arg Arg Ser Val Lys Ser Gly Pro Arg Glu Val Pro Arg
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Asp Glu Tyr Glu Asp Leu Tyr Tyr Thr Pro Ser Ser Gly Met Ala Ser
 20 25 30

Pro Asp Ser Pro Pro Asp Thr Ser Arg Arg Gly Ala Leu Gln Thr Arg
 35 40 45

Ser Arg Gln Arg Gly Glu Val Arg Phe Val Gln Tyr Asp Glu Ser Asp
 50 55 60

Tyr Ala Leu Tyr Gly Gly Ser Ser Ser Glu Asp Asp Glu His Pro Glu
 65 70 75 80

Val Pro Arg Thr Arg Arg Pro Val Ser Gly Ala Val Leu Ser Gly Pro
 85 90 95

Gly Pro Ala Arg Ala Pro Pro Pro Pro Ala Gly Ser Gly Gly Ala Gly
 100 105 110

Arg Thr Pro Thr Thr Ala Pro Arg Ala Pro Arg Thr Gln Arg Val Ala
 115 120 125

Ser Lys Ala Pro Ala Ala Pro Ala Ala Glu Thr Thr Arg Gly Arg Lys
 130 135 140

59

Ser Ala Gln Pro Glu Ser Ala Ala Leu Pro Asp Ala Pro Ala Ser Thr
 145 150 155 160
 Ala Pro Thr Arg Ser Lys Thr Pro Ala Gln Gly Leu Ala Arg Lys Leu
 165 170 175
 His Phe Ser Thr Ala Pro Pro Asn Pro Asp Ala Pro Trp Thr Pro Arg
 180 185 190
 Val Ala Gly Phe Asn Lys Arg Val Phe Cys Ala Ala Val Gly Arg Leu
 195 200 205
 Ala Ala Met His Ala Arg Met Ala Ala Val Gln Leu Trp Asp Met Ser
 210 215 220
 Arg Pro Arg Thr Asp Glu Asp Leu Asn Glu Leu Leu Gly Ile Thr Thr
 225 230 235 240
 Ile Arg Val Thr Val Cys Glu Gly Lys Asn Leu Leu Gln Arg Ala Asn
 245 250 255
 Glu Leu Val Asn Pro Asp Val Val Gln Asp Val Asp Ala Ala Thr Ala
 260 265 270
 Thr Arg Gly Arg Ser Ala Ala Ser Arg Pro Thr Glu Arg Pro Arg Ala
 275 280 285
 Pro Ala Arg Ser Ala Ser Arg Pro Arg Arg Pro Val Glu Gly Thr Glu
 290 295 300
 Leu Gly Ser Thr Ser Pro Val Trp Trp Asn Ser Ala Asp Ile Gln His
 305 310 315 320
 Ser Gly Gly Arg Met Ser Asn Leu Leu Thr Val His Gln Asn Leu Pro
 325 330 335
 Ala Leu Pro Val Asp Ala Thr Ser Asp Glu Val Arg Lys Asn Leu Met
 340 345 350
 Asp Met Phe Arg Asp Arg Gln Ala Phe Ser Glu His Thr Trp Lys Met
 355 360 365
 Leu Leu Ser Val Cys Arg Ser Trp Ala Ala Trp Cys Lys Leu Asn Asn
 370 375 380
 Arg Lys Trp Phe Pro Ala Glu Pro Glu Asp Val Arg Asp Tyr Leu Leu
 385 390 395 400
 Tyr Leu Gln Ala Arg Gly Leu Ala Val Lys Thr Ile Gln Gln His Leu
 405 410 415
 Gly Gln Leu Asn Met Leu His Arg Arg Ser Gly Leu Pro Arg Pro Ser
 420 425 430
 Asp Ser Asn Ala Val Ser Leu Val Met Arg Arg Ile Arg Lys Glu Asn
 435 440 445
 Val Asp Ala Gly Glu Arg Ala Lys Gln Ala Leu Ala Phe Glu Arg Thr
 450 455 460
 Asp Phe Asp Gln Val Arg Ser Leu Met Glu Asn Ser Asp Arg Cys Gln
 465 470 475 480

60

Asp Ile Arg Asn Leu Ala Phe Leu Gly Ile Ala Tyr Asn Thr Leu Leu
 485 490 495
 Arg Ile Ala Glu Ile Ala Arg Ile Arg Val Lys Asp Ile Ser Arg Thr
 500 505 510
 Asp Gly Gly Arg Met Leu Ile His Ile Gly Arg Thr Lys Thr Leu Val
 515 520 525
 Ser Thr Ala Gly Val Glu Lys Ala Leu Ser Leu Gly Val Thr Lys Leu
 530 535 540
 Val Glu Arg Trp Ile Ser Val Ser Gly Val Ala Asp Asp Pro Asn Asn
 545 550 555 560
 Tyr Leu Phe Cys Arg Val Arg Lys Asn Gly Val Ala Ala Pro Ser Ala
 565 570 575
 Thr Ser Gln Leu Ser Thr Arg Ala Leu Glu Gly Ile Phe Glu Ala Thr
 580 585 590
 His Arg Leu Ile Tyr Gly Ala Lys Asp Asp Ser Gly Gln Arg Tyr Leu
 595 600 605
 Ala Trp Ser Gly His Ser Ala Arg Val Gly Ala Ala Arg Asp Met Ala
 610 615 620
 Arg Ala Gly Val Ser Ile Pro Glu Ile Met Gln Ala Gly Gly Trp Thr
 625 630 635 640
 Asn Val Asn Ile Val Met Asn Tyr Ile Arg Asn Leu Asp Ser Glu Thr
 645 650 655
 Gly Ala Met Val Arg Leu Leu Glu Asp Gly Asp Gly Ile Glu Gly Arg
 660 665 670
 Gly Ser Ala Trp Arg His Pro Gln Phe Gly Gly
 675 680

<210> 22

<211> 11

<212> PRT

<213> Artificial Sequence

<220>

 <223> Description of Artificial Sequence:synthetic TAT
 protein

<400> 22

 Ala Gly Arg Lys Lys Arg Arg Gln Arg Arg Arg
 1 5 10

<210> 23

<211> 11

<212> PRT

<213> Artificial Sequence

<220>

 <223> Description of Artificial Sequence: synthetic TAT
 protein

<400> 23

Tyr Ala Arg Lys Ala Arg Arg Gln Ala Arg Arg
1 5 10

<210> 24

<211> 11

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic TAT
protein

<400> 24

Tyr Ala Arg Ala Ala Ala Arg Gln Ala Arg Ala
1 5 10

<210> 25

<211> 11

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic TAT
protein

<400> 25

Tyr Ala Arg Ala Ala Arg Arg Ala Ala Arg Arg
1 5 10

<210> 26

<211> 11

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic TAT
protein

<400> 26

Tyr Ala Arg Ala Ala Arg Arg Ala Ala Arg Ala
1 5 10

<210> 27

<211> 11

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic TAT
protein

<400> 27

Tyr Ala Arg Arg Arg Arg Arg Arg Arg Arg
1 5 10

<210> 28

<211> 11

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic TAT protein

<400> 28

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1 5 10

<210> 29

<211> 4960

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: vector pCMV-I-Cre-pA

<400> 29

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Claims

1. Use of a fusion protein comprising
 - (a) a site-specific DNA recombinase domain and
 - (b) a protein transduction domain (PTD)for preparing an agent for inducing target gene alterations in a living organism or cell culture, wherein said living organism carries at least one or more recognition sites for said site-specific DNA recombinase integrated in an endogenous gene.
2. The use of claim 1, wherein the PTD is not derived from Antennapedia and preferably is a PTD derived from the VP22 protein of HSV or from the TAT protein of HIV.
3. Use of a fusion protein comprising
 - (a) a site-specific DNA recombinase domain and
 - (b) a protein transduction domain (PTD) being not derived from Antennapedia and preferably being derived from the VP22 protein of HSV or from the TAT protein of HIVfor preparing an agent for inducing target gene alterations in a living organism or cell culture, wherein said living organism carries at least one or more recognition sites for said site-specific DNA recombinase integrated in its genome.
4. The use of claim 3, wherein the recognition sites for said site specific recombinase is present within an endogenous gene or a transgene.
5. The use of any one of claims 2 to 4, wherein the TAT protein comprises
 - (i) the amino acid sequence YGRKKRRQRRR (SEQ ID NO: 10) or a mutant thereof including
 - (ii) peptides having the amino sequences

AGRKKRRQRRR (SEQ ID NO:22)

YARKARRQARR (SEQ ID NO:23)

YARAAARQARA (SEQ ID NO:24)

YARAARRAARR (SEQ ID NO:25)

YARAARRAARA (SEQ ID NO:26)

YARRRRRRRRR (SEQ ID NO:27)

YAAARRRRRRR (SEQ ID NO:28);

preferably the TAT protein consists of one of the sequences shown in (i) or (ii) above.

6. The use of any one of claims 2 to 4, wherein the VP22 protein comprises the amino acid 16-157 of SEQ ID NO:14.

7. The use of any one of claims 1 to 6, wherein the site-specific DNA recombinase domain is selected from a recombinase protein derived from Cre, Flp, ϕ C31 recombinase, and R recombinase and preferably is Cre having amino acids 15 to 357 of SEQ ID NO: 2 or Flpe having amino acids 15 to 437 of SEQ ID NO: 4.

8. The use of any one of claims 1 to 7, wherein the protein transduction domain is fused to the N-terminal of the site-specific DNA recombinase domain.

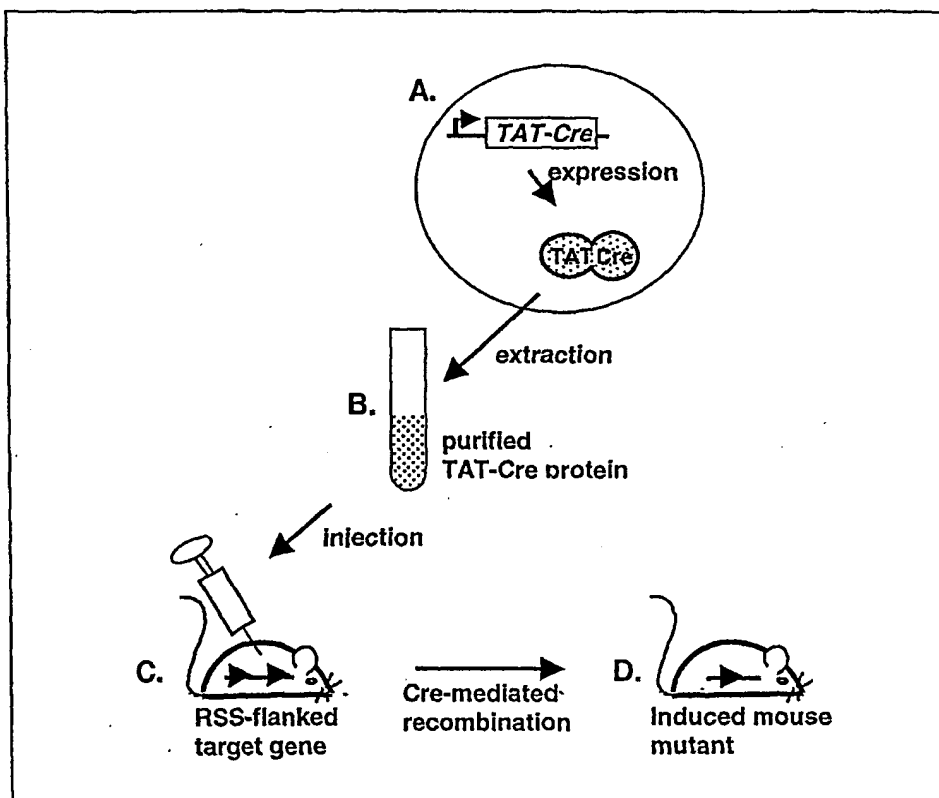
9. The use of any one of claims 1 to 8, wherein the protein transduction domain is fused to the site-specific DNA recombinase domain through a direct chemical bond or through a linker molecule.

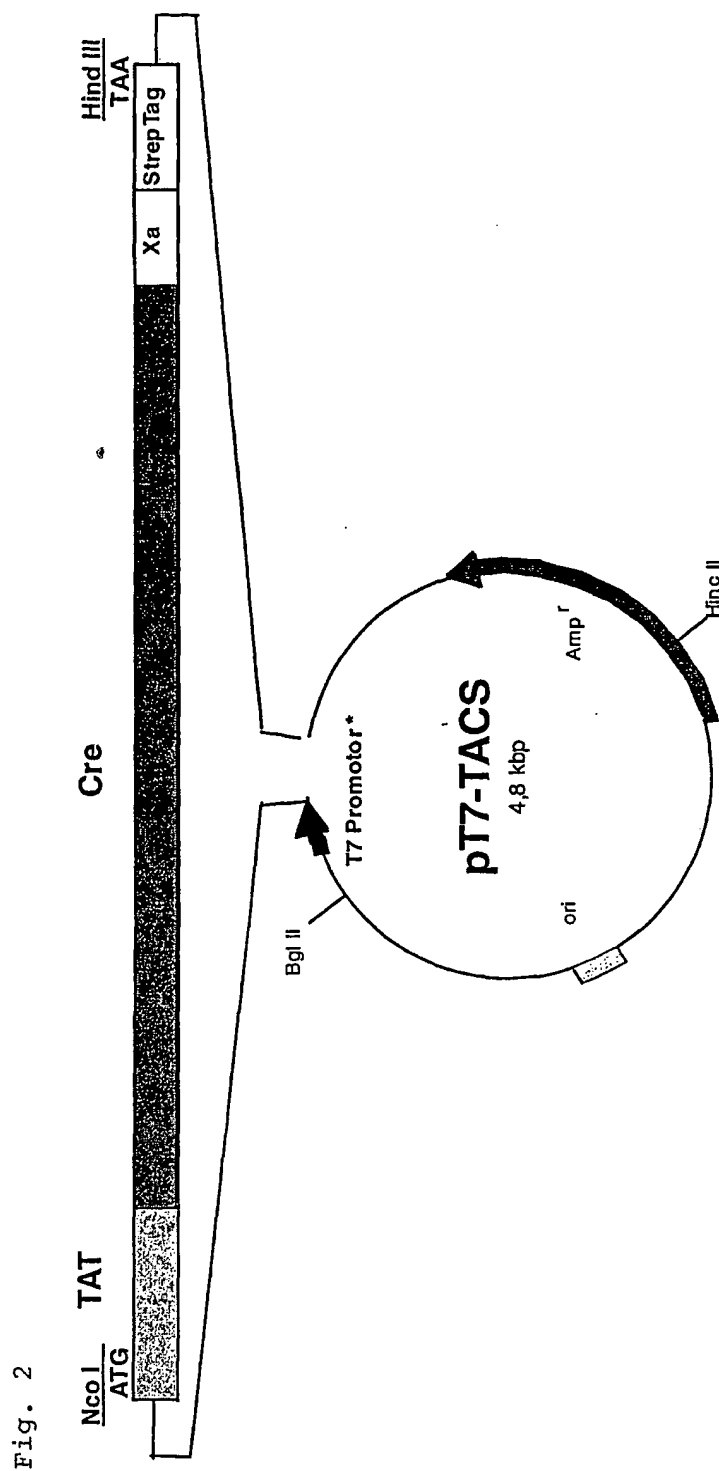
10. The use of any one of claim 9, wherein the linker molecule is a short peptide having 1 to 20, preferably 1 to 10 amino acid residues.

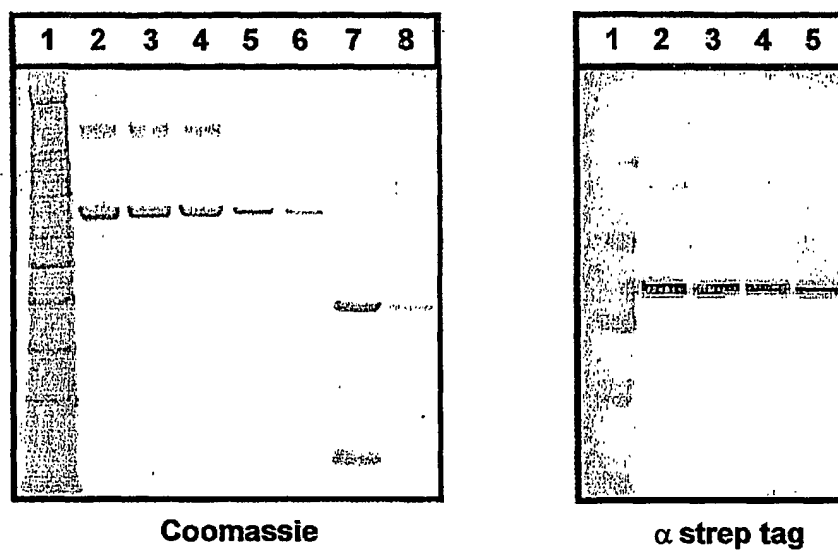
11. The use of any one of claims 1 to 10, wherein said fusion protein further comprises additional functional sequences.
12. The use of claim 1, wherein the fusion protein has the sequence shown in SEQ ID NOs: 2, 4, 6 or 8.
13. The use of any one of claims 1 to 12, wherein the living organism is a vertebrate, preferably a rodent or a fish.
14. A method for inducing gene alterations in a living organism which comprises administering to said living organism, a fusion protein comprising a site-specific DNA recombinase domain and a protein transduction domain as defined in claims 1 to 12, wherein said living organism carries at least one or more recognition sites for said site-specific DNA recombinase *integrated in its genome*.
15. A fusion protein comprising
 - (a) a site-specific DNA recombinase domain as defined in claims 2 to 9 and
 - (b) a protein transduction domain (PTD) as defined in claims 2 to 9 provided that when (a) is the wild-type Flp or Cre then (b) is not the full length VP22 protein of HSV.
16. The fusion of claim 15, wherein the (PTD) is derived from the TAT protein of HIV.
17. A DNA sequence coding for the fusion protein of claim 15 or 16, said DNA sequence preferably comprising the sequence shown in SEQ ID NOs: 1, 3, 5, 7, 9, 11, 13, 18 and/or 20.
18. A vector comprising the DNA sequence of claim 17.

19. A host cell transformed with the vector of claim 18 and/or comprising the DNA of claim 17.
20. A method for producing the fusion protein of claim 15 which comprises culturing the transformed host cell of claim 19 and Isolating the fusion protein.
21. An injectable composition comprising the fusion protein as defined in claims 1 to 12 or 15 to 16.

Fig. 1





**Figure 3**

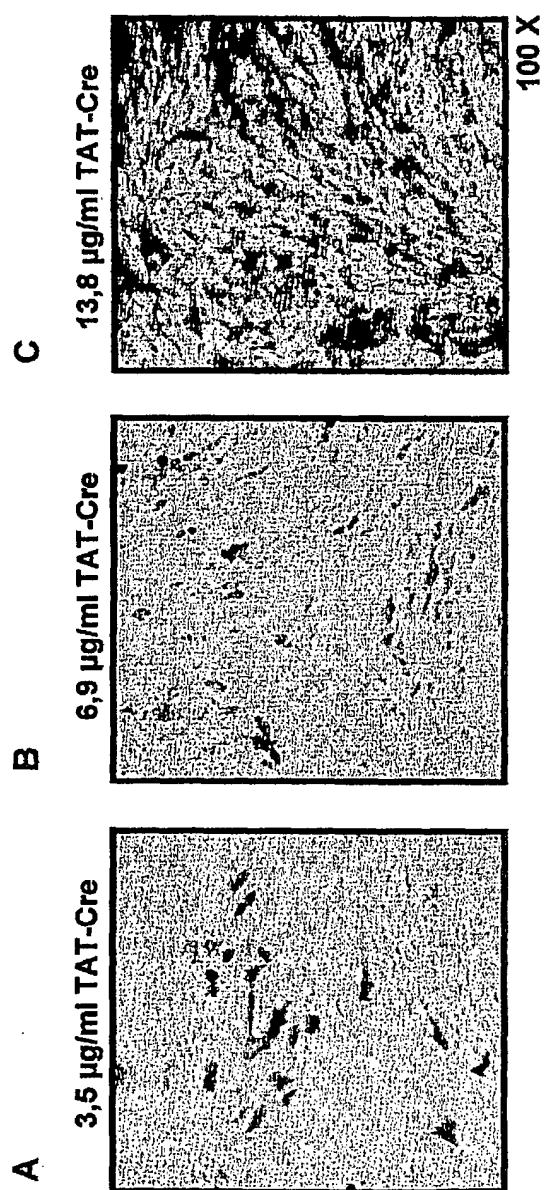
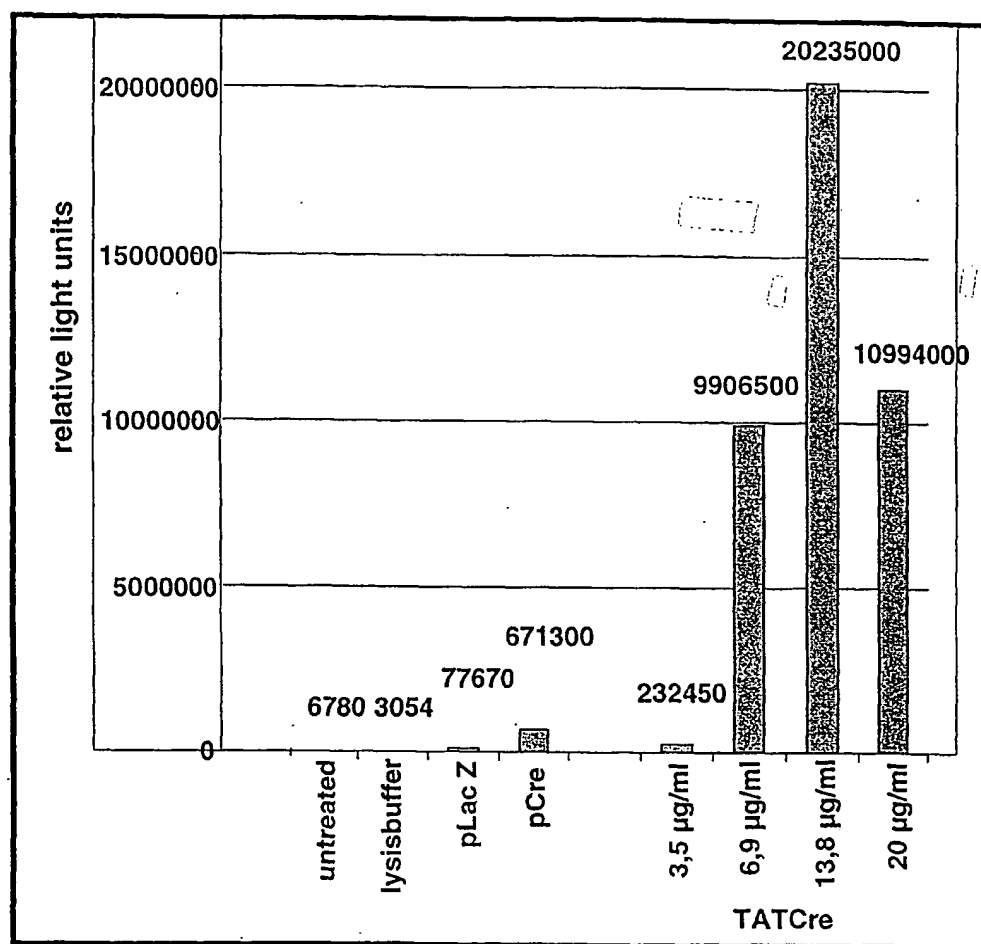
**Figure 4**

Fig. 5



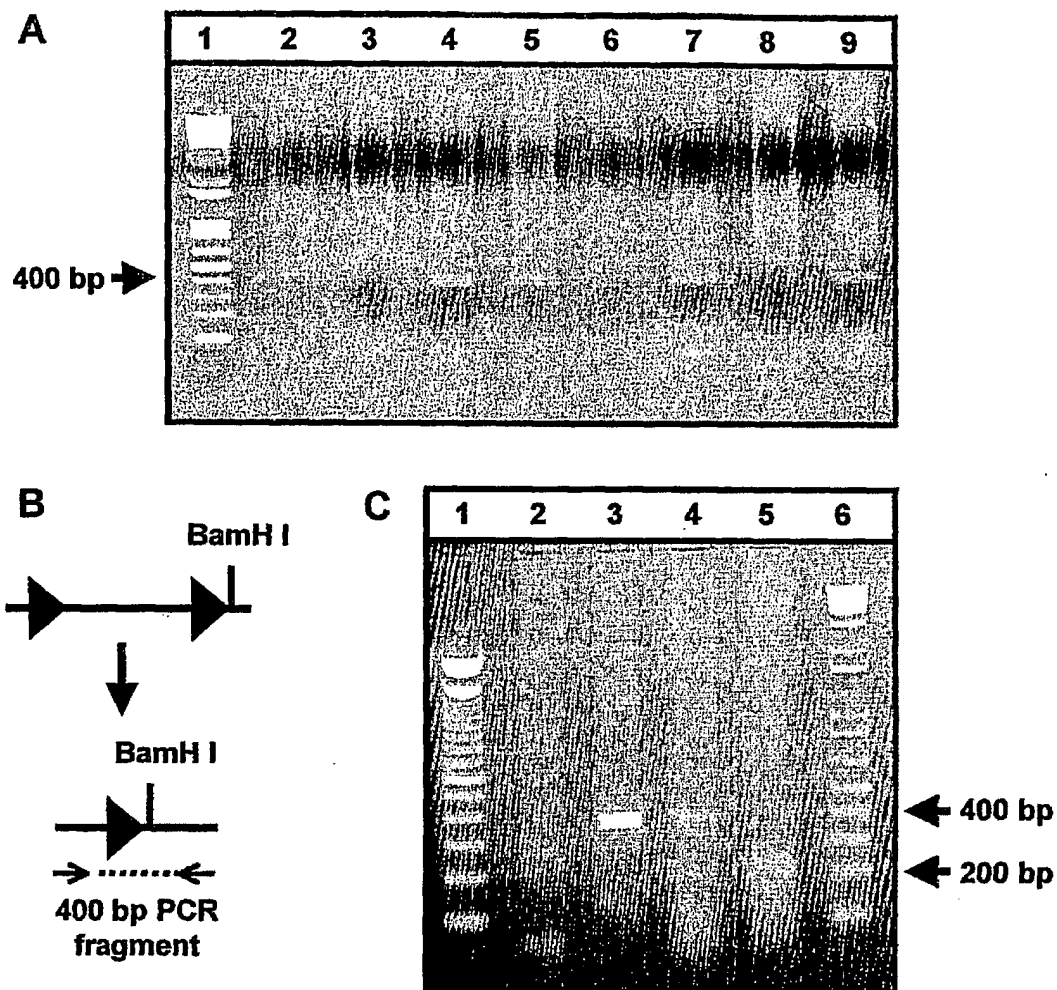
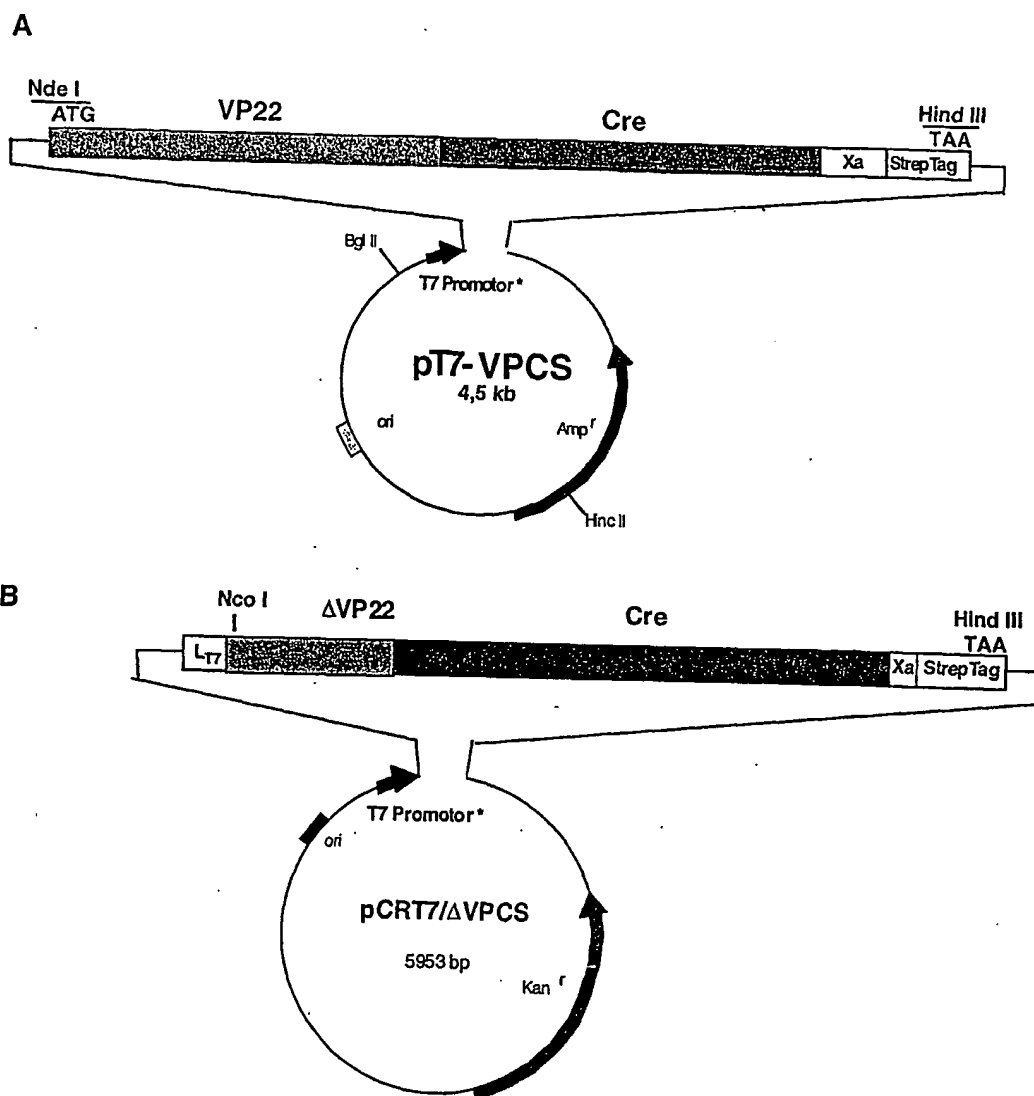


Figure 6

Fig. 7



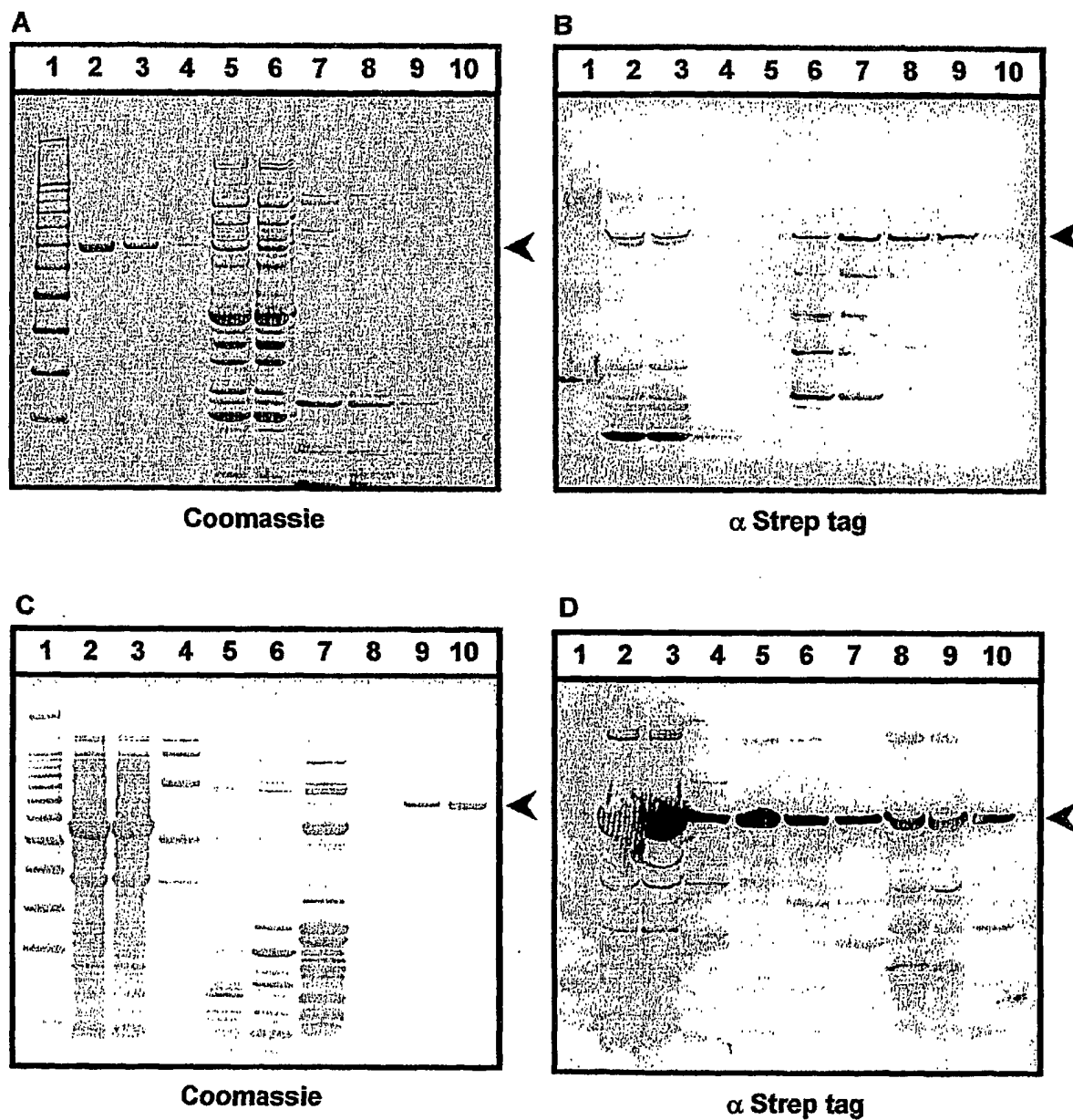


Figure 8

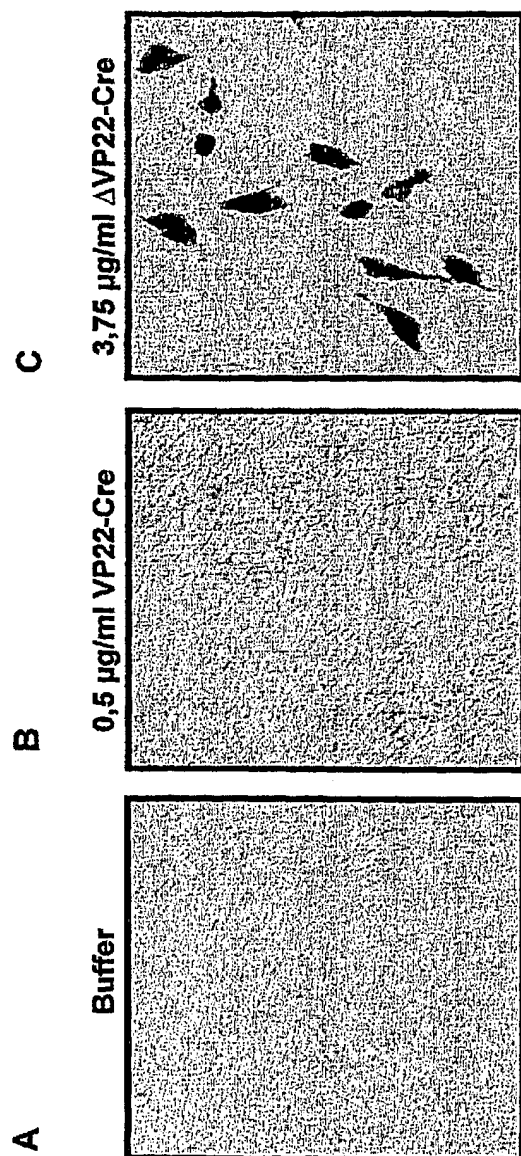
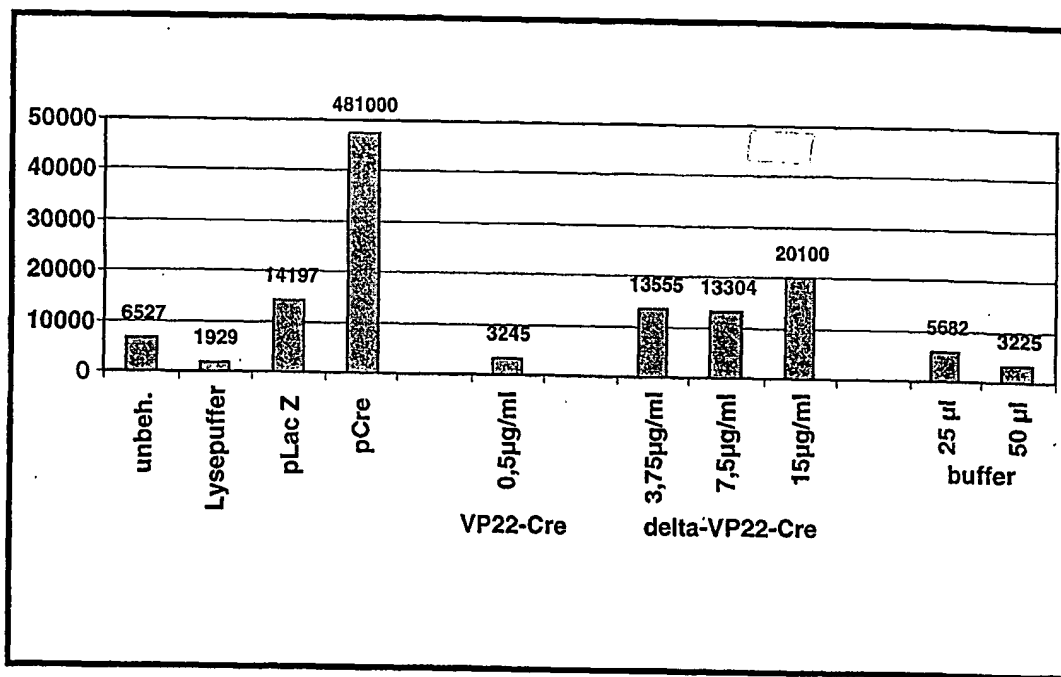


Figure 9

Fig. 10



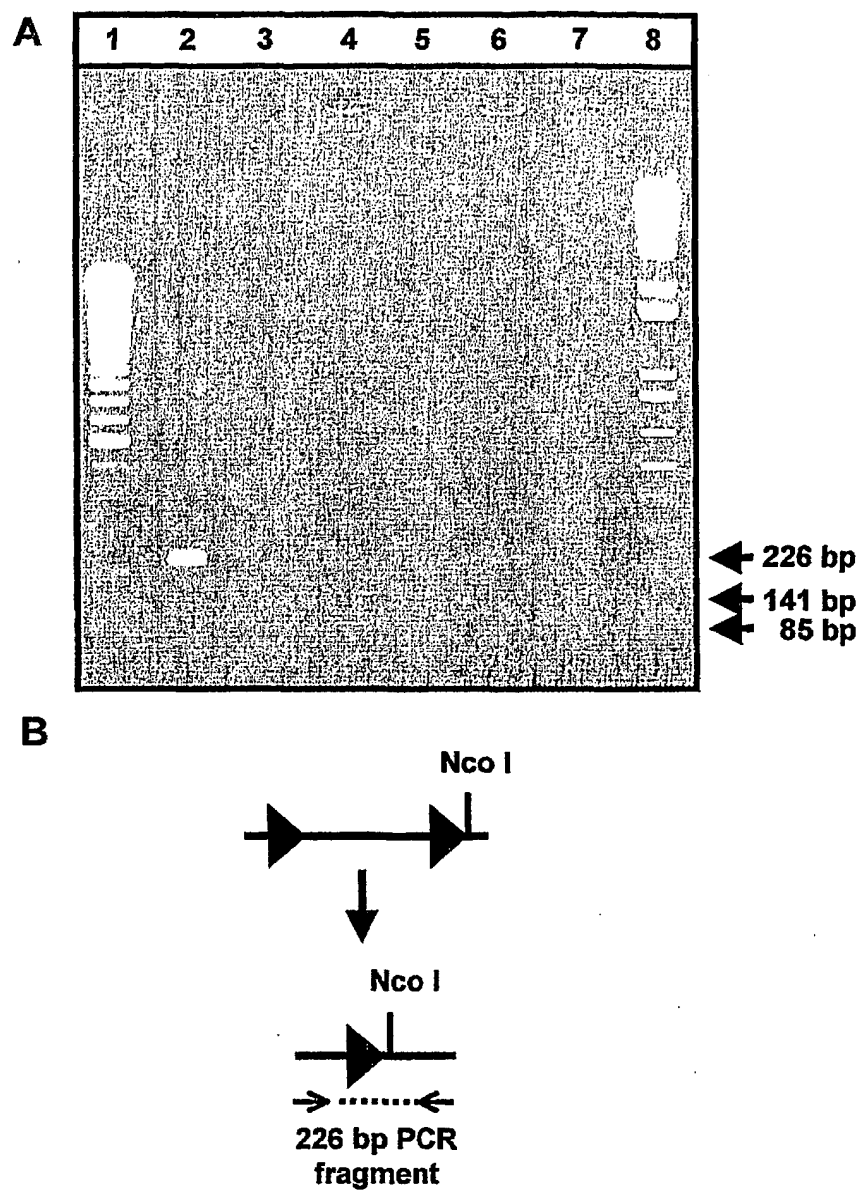


Figure 11

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